

Award Number: DAMD17-01-1-0172

TITLE: A Novel Molecular Target for Breast Cancer Prevention and Treatment

PRINCIPAL INVESTIGATOR: Xiao-Kun Zhang, Ph.D.

CONTRACTING ORGANIZATION: The Burnham Institute
La Jolla, California 92037

REPORT DATE: June 2003

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20040226 029

REPORT DOCUMENTATION PAGE

Form Approved
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE June 2003	3. REPORT TYPE AND DATES COVERED Annual (1 Jun 2002 - 31 May 2003)
4. TITLE AND SUBTITLE A Novel Molecular Target for Breast Cancer Prevention and Treatment			5. FUNDING NUMBERS DAMD17-01-1-0172
6. AUTHOR(S) Xiao-Kun Zhang, Ph.D.			
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) The Burnham Institute La Jolla, California 92037 E-Mail: xzhang@burnham.org			8. PERFORMING ORGANIZATION REPORT NUMBER
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER
11. SUPPLEMENTARY NOTES Original contains color plates: All DTIC reproductions will be in black and white.			
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited			12b. DISTRIBUTION CODE
13. ABSTRACT (Maximum 200 Words) Induction of apoptosis is a plausible approach for developing new cancer therapies. A class of synthetic retinoids related to 6-[3-(1-adamantlyl)-4-hydroxyphenyl]-2-naphthalene carboxylic acid (AHPN/CD437) potentially induce apoptosis of both hormone-dependent and -independent breast cancers. We previously reported that apoptotic effects of AHPN/CD437 are mediated by mitochondrial targeting of orphan nuclear receptor TR3. In the past year, we have studied the mechanisms by which TR3 migrates from the nucleus to the cytoplasm and by which TR3 targets mitochondria. Our results demonstrate that the migration of TR3 from the nucleus to the cytoplasm requires retinoid X receptor (RXR) through their unique heterodimerization and is regulated by RXR ligands. We have also identified a nuclear export sequence in the RXR, which is required for its cytoplasmic localization. Furthermore, we have found that TR3 targets mitochondria through its interaction with Bcl-2 that resides at the outer mitochondrial membrane. The interaction converts Bcl-2 from an anti-apoptotic to a pro-apoptotic molecule. Our results not only enhance our understanding of the molecular mechanism by which TR3 exerts its apoptotic effects in breast cancer cells but also provide novel approaches to induce apoptosis of Bcl-2-expressing breast cancer cells by using RXR ligands.			
14. SUBJECT TERMS No Subject Terms Provided.			15. NUMBER OF PAGES 34
			16. PRICE CODE
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89)
Prescribed by ANSI Std. Z39-18
298-102

Table of Contents

Cover	1
SF 298	2
Table of Contents	3
Introduction	4
Body	4
Key Research Accomplishments	4
Reportable Outcomes	15
Conclusions	15
References	16
Appendices	17

INTRODUCTION

Although anti-estrogen tamoxifen shows activities in preventing development of breast cancer, there are no effective therapies for malignant estrogen-independent breast cancer, which is also often resistant to chemotherapy and γ -radiation therapy. Lack of such therapies has contributed to the high mortality for U.S. women. Novel therapies are therefore urgently needed.

Recent scientific progress has suggested that elimination of cancer cells through induction of cancer cell death (apoptosis) is a plausible approach for developing new cancer therapies. Retinoids, analogs of vitamin A, are promising chemopreventive agents for breast cancer. Recently, a new class of synthetic retinoids related to 6-[3-(1-adamantyl)-4-hydroxyphenyl]-2-naphthalene carboxylic acid (AHPN/CD437) were found to potently induce apoptosis of both hormone-dependent and -independent breast cancers (1), indicating that it may be representative of a novel class of compounds suitable for treatment of estrogen-independent breast cancers. We subsequently showed that TR3, an orphan member of the steroid/thyroid/retinoid receptor superfamily, is required for induction of apoptosis of lung and prostate cancer cells by AHPN/CD437 and other apoptotic stimuli (2). Moreover, we discovered that TR3, in response to apoptosis stimuli, is translocated from the nucleus to the cytoplasm, where it targets mitochondria to induce cytochrome c release and apoptosis (3, 4).

The objective of this application is to develop evidence that TR3 is a suitable molecular target for developing drugs against breast cancer. In the proposed studies, we will first establish whether TR3 is required for proliferation and apoptosis of breast cancer cells in response to various stimuli. We will then determine whether the novel nuclear-to-mitochondrial pathway for apoptosis occurs in breast cancer cells. In addition, we will study mechanism by which translocation of TR3 from the nucleus to cytoplasm is regulated by studying effect of phosphorylation. Furthermore, we will study physical interactions between TR3 and Bcl-2 and its effect on Bcl-2-mediated mitochondrial and post-mitochondrial activities. In the first year of research, we showed that retinoids AHPN analogs SR11453 and MM002 potently induced TR3 expression and apoptosis in breast cancer cells. Furthermore, we found that AHPN analogs induced migration of TR3 from the nucleus to mitochondria. We also observed that mitochondrial targeting of TR3 was regulated by ligands for retinoid X receptor (RXR) and that TR3 interacted with Bcl-2. In the past year, we have continued our studies on these directions and have made substantial progress as described below.

KEY RESEARCH ACCOMPLISHMENTS

1. Regulation of TR3 nuclear export by retinoid X receptor and its ligands.

Mitochondrial localization of TR3 in breast cancer cells. Fontana's group first showed that a new class of synthetic retinoids, related to 6-[3-(1-adamantyl)-4-hydroxyphenyl]-2-naphthalene carboxylic acid (AHPN/CD437) effectively induced apoptosis of both estrogen-dependent and -independent breast cancer cells (1), suggesting that these retinoids may represent a new class of drugs that have therapeutic value for the treatment of estrogen-independent breast cancer. We recently demonstrated that retinoid AHPN-induced apoptosis required expression of orphan receptor TR3, which migrated from the nucleus to mitochondria to trigger apoptosis (3). In our first year of research funded by BCRP, we showed that a green fluorescent protein (GFP)-TR3 fusion transfected into MDA-MB-231 cells migrated from the nucleus to the cytoplasm where it targeted to mitochondria when MDA-MB-231 cells were treated with two AHPN analogs, SR11453 and SR11384 (Figure 1A). Furthermore, we observed that GFP-TR3/ Δ DBD (a mutant that lacks the DNA-binding domain) targeted mitochondria in MDA-MB-231 and ZR-75-1 breast cancer cells (Figure 1B).

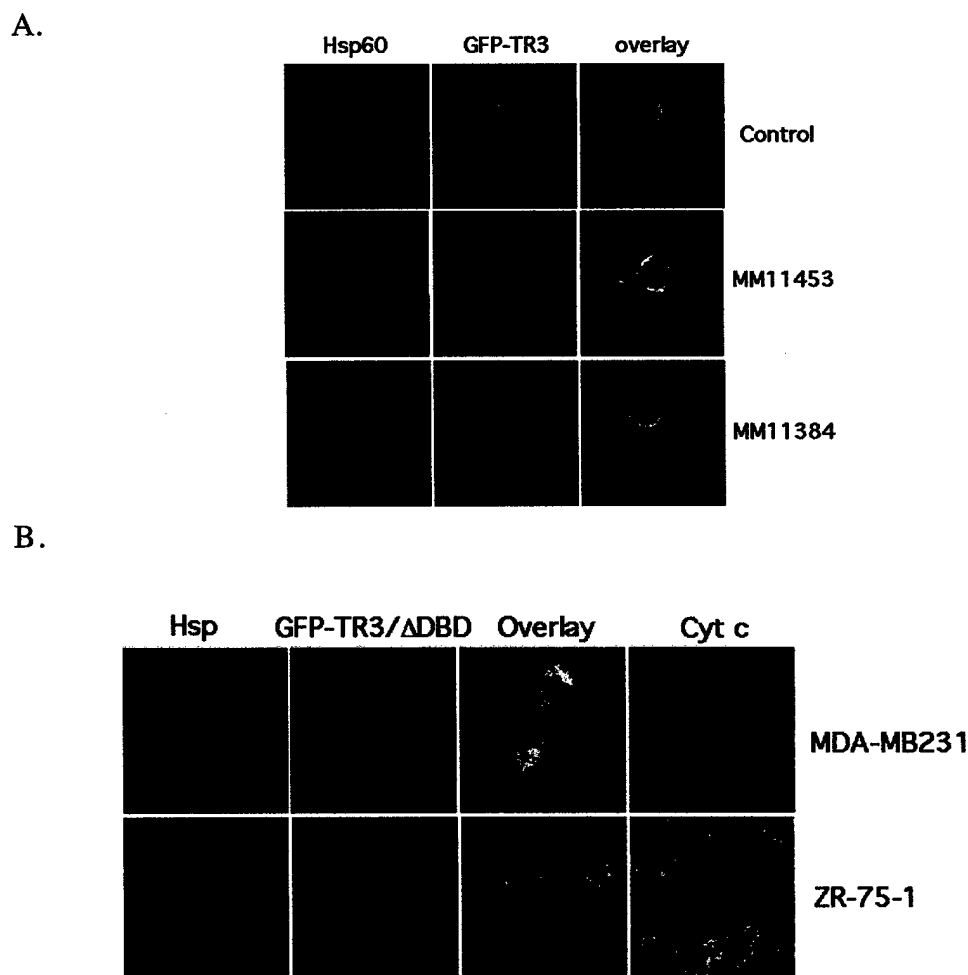


Figure 1. A. TR3 is targeted to mitochondria in MDA-MB-231 cells in response to an apoptosis inducer. GFP-TR3-transfected MDA-MB-231 cells were treated with or without SR11453 (10^{-6} M) or SR11384 (10^{-6} M) for 1 h, then immunostained with anti-Hsp60 antibody (Sigma), followed by Cy3-conjugated secondary antibody (Sigma) to detect mitochondria. GFP-TR3 and mitochondria (Hsp60) were visualized using confocal microscopy, and the two images were overlaid (see Overlay). **B. Mitochondrial targeting of TR3/ Δ DBD is associated with cytochrome c release.** GFP-TR3/ Δ DBD was transiently transfected into ZR-75-1 or MDA-MB-231 cells, which were stained for mitochondria (Hsp60) and cytochrome c (Cyt c), and analyzed by confocal microscopy.

RXR α targets mitochondria in response to apoptotic stimuli. How TR3 migrates from the nucleus to mitochondria to induce cytochrome c release is unknown. Since RXR α heterodimerizes with TR3 (5, 6), we studied whether RXR α targeted mitochondria in cells undergoing apoptosis. Subcellular localization of RXR α in LNCaP cells in the absence or presence of TPA, which potently induces LNCaP cell apoptosis, was examined by confocal microscopy analysis. Immunostaining showed that RXR α predominantly localized in the nucleus in the absence of TPA treatment (Figure 2A). However, when cells were treated with TPA, RXR α was found in the cytoplasm. To study whether RXR α was associated with mitochondria, cells were stained for heat shock protein 60 (Hsp60), a mitochondria-specific protein (Figure 2A). The extensive overlap in the distribution patterns of Hsp60 and RXR α suggested the association of RXR α with mitochondria. Similarly, treatment with TPA also resulted in mitochondrial localization of TR3 (Figure 2B), as previously reported (3).

RXR α and TR3 mitochondrial targeting are mutually dependent. To study whether TR3 and RXR α targeted mitochondria as a heterodimer, subcellular localization of TR3 and RXR α was examined in LNCaP cells treated with or without TPA. In the absence of TPA, both TR3 and RXR α resided mainly in the nucleus (Figure 2C). However, when cells were treated with TPA, TR3 and RXR α were colocalized in the cytoplasm and their distribution patterns overlaid extensively (Figure 2C), suggesting their association in the cytoplasm. Furthermore, transfected RXR α and TR3 associated and targeted mitochondria when cells were treated with an apoptosis inducer, the AHPN analog 3-CI-AHPC (also called MM002) (7) that potently induced the apoptosis of cancer cells (Figure 2D). We next determined whether RXR α cytoplasmic localization depended on TR3 expression by examining its subcellular localization in LNCaP cells stably expressing TR3 antisense RNA (3). Expression of TR3 antisense RNA strongly inhibited TPA-induced TR3 expression in LNCaP cells. In contrast to that observed in wild-type LNCaP cells (Figure 2A), RXR α was found only in the nucleus in the TR3 antisense stable clone, even though the cells were treated with TPA (Figure 2E). To study whether TR3 mitochondrial targeting required RXR α , we used the siRNA approach to inhibit RXR α expression in LNCaP cells, then examined the subcellular localization of TR3. Transfection of LNCaP cells with RXR α siRNA strongly reduced RXR α protein levels (Figure 2F). In cells transfected with RXR α siRNA, TR3 was mainly confined in the nucleus despite TPA treatment (Figure 2F). Thus, cytoplasmic localization of TR3 and RXR α is mutually dependent.

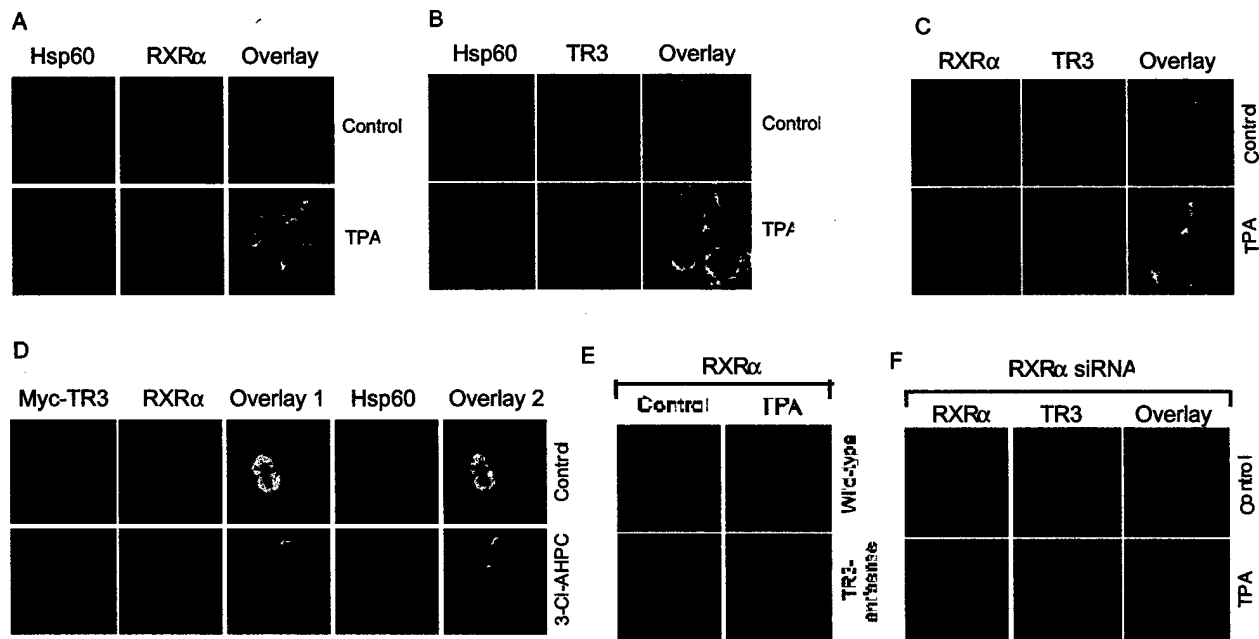


Figure 2. TR3 and RXR comigrate from the nucleus to the cytoplasm in LNCaP cells. RXR α (A) or TR3 (B) targets mitochondria in response to apoptotic stimulus. LNCaP cells were treated with TPA (100 ng/ml) for 1 hr, then immunostained with either anti-RXR α (Santa Cruz) (A) or anti-TR3 (B) antibody to detect RXR α or TR3 or with anti-Hsp60 antibody to detect mitochondria. RXR α , TR3 and mitochondria (Hsp60) were visualized using confocal microscopy, and the images of RXR α or TR3 were overlaid with those of mitochondria (overlay). **(C) TR3 and RXR α comigrate from the nucleus to the cytoplasm.** LNCaP cells were treated with or without TPA for 1 hr, then immunostained with anti-RXR α or anti-TR3 antibody. RXR α and TR3 were visualized using confocal microscopy and the images were overlaid (overlay). **(D) 3-CI-AHPC induces mitochondrial localization of transfected RXR α and TR3.** Expression vectors for myc-TR3 and RXR α were transfected into LNCaP cells. Cells

were treated with 3-Cl-AHPC for 3 hr, then immunostained with anti-myc or anti-RXR α antibody. Myc-TR3, RXR α , and Hsp60 were visualized and the images were overlaid. In Overlay1, the myc-TR3 and RXR α images are overlaid, and in Overlay2 the myc-TR3, RXR α , and Hsp60 images are overlaid. **(E) Mitochondrial localization of RXR α is TR3 dependent.** LNCaP cells or LNCaP cells stably expressing TR3 antisense RNA (TR3/Antisense) were treated with or without TPA for 1 hr, then immunostained with anti-RXR α antibody. **(F) Effect of RXR siRNA on TR3 localization.** LNCaP cells were transfected with or without RXR α siRNA or control siRNA for 72 hr. Cells were analyzed for subcellular localization of RXR α and TR3 by confocal microscopy.

Identification of a putative NES in RXR α . To study whether the cytoplasmic localization of the RXR α /TR3 heterodimer is mediated through CRM1-dependent nuclear export we examined the effect of the inhibitor leptomycin B (LMB) on this type of export. Treatment of LNCaP cells with LMB completely blocked TPA-induced cytoplasmic localization of TR3 and RXR α (Figure 3A). This result suggests that the nuclear export of RXR α /TR3 heterodimer is mediated by the CRM1-dependent mechanism.

CRM1-dependent nuclear export is mediated by a Leucine-rich nuclear export signal (NES). Our mutagenesis studies revealed the presence of a Methionine-rich sequence, which has significant homology to various identified NESs (Figure 3B). The DNA sequences representing RXR α residues 348-368 were fused to green fluorescent protein (GFP), and the fusion was transfected into HEK293T embryonic kidney cells. HEK293T cells were chosen for our mechanistic studies because of their high transfection efficiency. The GFP-NES fusion was found exclusively in the cytoplasm, while LMB treatment prevented its nuclear export. Mutations of Met357 and Met360 to Ala (GFP-NESm-RXR α) largely abolished nuclear export activity (Figure 3C). Thus, RXR α amino acids 348-368 represent a putative NES.

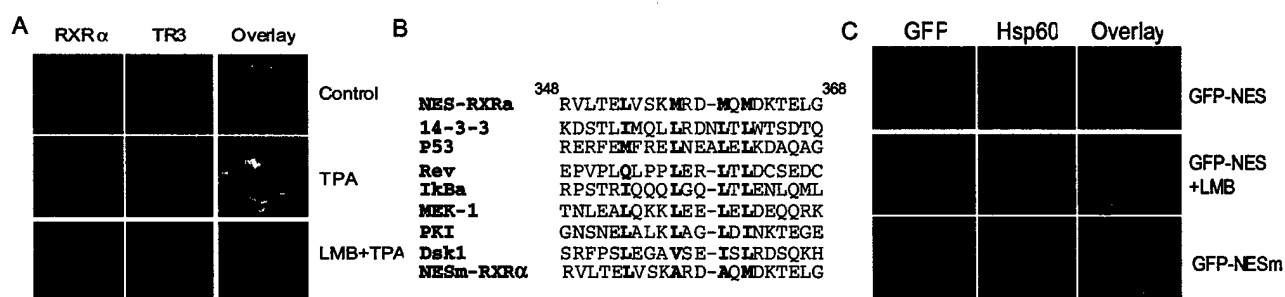


Figure 3. Identification of a nuclear export sequence in RXR α . **(A)** Cytoplasmic localization of RXR α /TR3 is mediated by CRM1-dependent nuclear export. LNCaP cells were treated with TPA in the absence (control) or presence of leptomycin B (LMB, 2.5 ng/ml) (Sigma) and analyzed by confocal microscopy. **(B)** Schematic representation of the RXR α NES. The identified RXR α NES is compared with known NESs identified in the indicated genes. The bold letters indicate conserved amino acid residues. **(C)** The RXR α NES is capable of directing GFP to the cytoplasm. The putative RXR α NES (RVLTELVS~~KMRD~~MQMDKTELG) or its mutant (RVLTELVS~~KARD~~AQMDKTELG) (NESm) was fused to GFP, and the expression vectors were transfected into HEK293T embryonic kidney cells. Cells were treated with or without LMB for 6 hr, then stained for Hsp60 and analyzed by confocal microscopy.

Regulation of the RXR α NES activity by ligand binding. To determine whether RXR α ligands inhibited RXR α NES activity through their induction of RXR α dimerization, we examined their effects on the cytoplasmic accumulation of RXR α /C1, a mutant RXR α ligand-binding domain that exclusively localized in the cytoplasm of HEK293T cells (data not shown). Treatment of GFP-RXR α /C1-transfected cells with RXR α ligand 9-*cis*-RA or SR11237 resulted in the diffuse distribution of RXR α /C1 throughout the cells, whereas treatment the RAR α subtype-selective Am80 produced no such effect (Figure 4A). Nondenaturing

polyacrylamide gel electrophoresis (PAGE) indicated that GFP-RXR α /C1 occurred only as a monomer in the cytoplasmic fraction of these cells. After treatment with 9-*cis*-RA, the monomeric form of GFP-RXR α /C1 in the cytoplasmic fraction disappeared, and only the homodimeric form was detected in the nuclear fraction of the lysate (Figure 4B). Thus, monomeric RXR α exists in the cytoplasm, while liganded-homodimeric RXR α is in the nucleus. RXR ligands 9-*cis*-RA and SR11237 inhibit RXR α nuclear export probably by inducing RXR α homodimerization.

We also analyzed the subcellular localization of two homodimerization defective-RXR α mutants, RXR α /385, which lacks the major homodimerization domain, and RXR α /LLL, which has Leu418, Leu420, and Leu430 in helix 10 replaced with Ala and fails to homodimerize in response to 9-*cis*-RA (8). Interestingly, both RXR α mutants were exclusively localized in the cytoplasm regardless of the presence of 9-*cis*-RA (Figure 4C). Another RXR α mutant, RXR α /C1/C432A, which also fails to homodimerize in response to 9-*cis*-RA (data not shown), was stained exclusively in the cytoplasm despite 9-*cis*-RA treatment (Figure 4C). Thus, subcellular localization of RXR α depends on its dimerization status and homodimerization suppresses RXR α nuclear export.

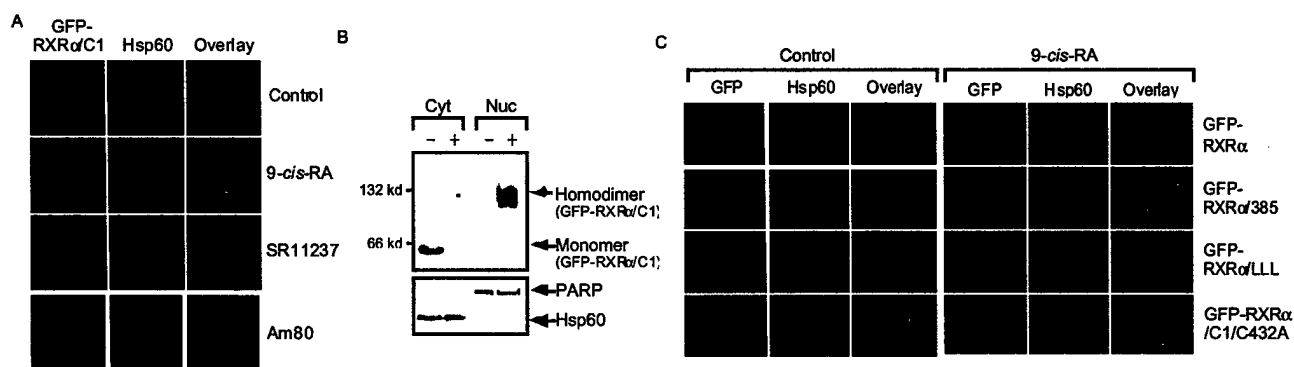


Figure 4. Regulation of RXR α nuclear export by its ligands and homodimerization. (A). Analysis of subcellular localization of RXR α /C1 in the absence or presence of retinoids. GFP-RXR α /C1 was transfected into HEK293T cells, which were then treated with the indicated retinoid, stained with Hsp60, and analyzed by confocal microscopy. **(B).** RXR α /C1 dimerization status determines its subcellular localization. GFP-RXR α /C1 was transfected into HEK293T cells, which were not treated or treated with 9-*cis*-RA (10^{-7} M). Nuclear and cytoplasmic extracts were prepared and analyzed by nondenaturing PAGE and anti-GFP antibody. The same extracts were analyzed by denaturing PAGE for expression of PARP and Hsp60 to ensure nuclear and mitochondrial fraction purity, respectively. **(C).** Confocal microscopy analysis of RXR homodimerization-defective mutants. GFP-RXR α /385, GFP-RXR α /LLL or GFP-RXR α /C1/C432A was transfected into HEK293T cells. Cells were treated with or without 9-*cis*-RA and analyzed by confocal microscopy.

Unique TR3/RXR α heterodimerization. Heterodimerization of RXR α with RAR α in solution largely depends on their dimerization interfaces localized in their LBDs and has been mapped to a region in the carboxyl (C)-terminus, corresponding to helices 9 and 10 in the canonical nuclear receptor LBD structure (9, 10). In GST pull-down assays, full-length RXR α was pulled-down by either GST-TR3 or GST-RAR α , whereas RXR α /385 was only pulled-down by GST-TR3 but not by GST-RAR α (Figure 5B). Thus, in solution the RXR α C-terminus is required for its interaction with RAR α but not with TR3. Similarly, as TR3/467, a mutant lacking the C-terminus, was effectively pulled-down by GST-RXR α (Figure 5C), the TR3 C-terminus was also dispensable for TR3 interaction with RXR α in solution. *In vivo* co-immunoprecipitation showed the efficient precipitation of GFP-RXR α /385 by anti-Flag antibody when Flag-TR3 was coexpressed in HEK293T cells (Figure 5D). In transactivation reporter gene assays, both RXR α /385 and RXR α similarly inhibited TR3 homodimer activity in the absence of 9-*cis*-RA (Figure 5E). Interestingly, TR3/467 and RXR α /385 colocalized in cells (Figure 5F). In addition to the LBD dimerization interface, RXR α contains another dimerization interface in the DNA-binding domain (DBD). Interestingly, TR3 also has a unique dimerization interface in its DBD called the C-terminal extension (11). Together, these results demonstrate that the C-termini of RXR α and TR3 are not required for their interaction in solution. Our results suggest that RXR α /TR3 heterodimerization in solution is mediated by the DBD dimerization interfaces (see also Figure 11). Such a unique heterodimeric structure may allow the activation of the RXR α NES.

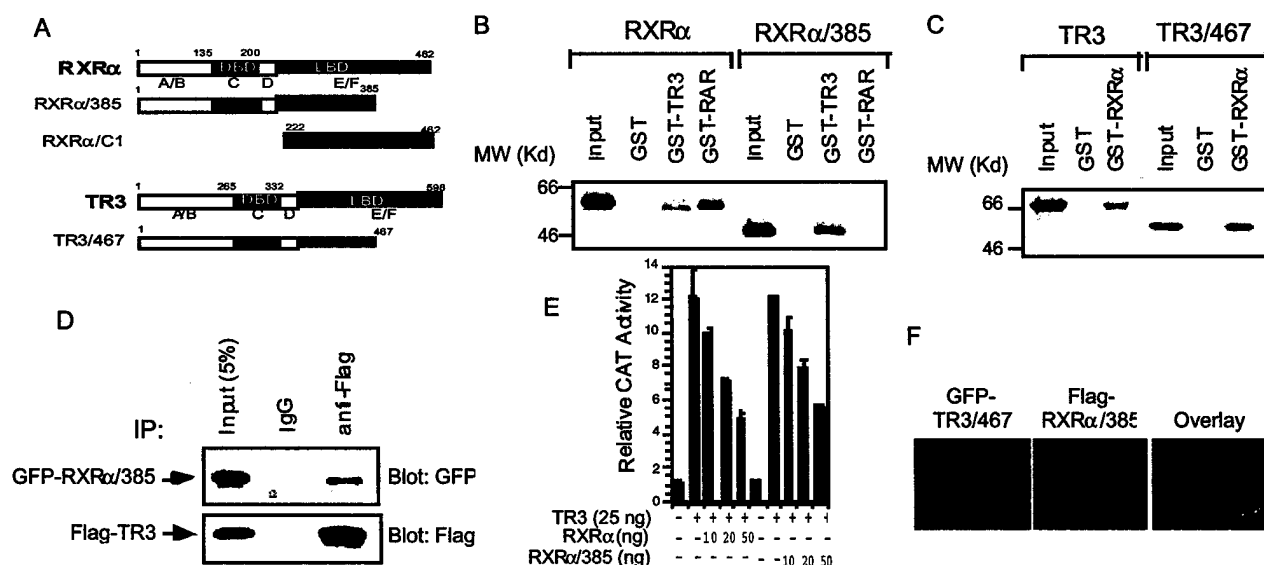
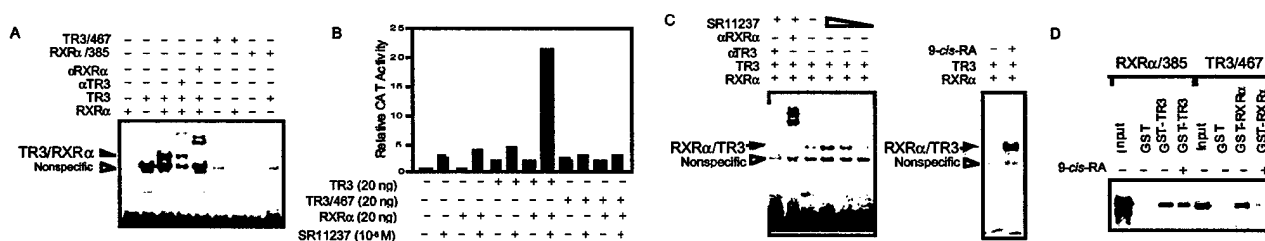


Figure 5. C-termini of RXR α and TR3 are not required for RXR α /TR3 interaction in solution. (A) Schematic representation of RXR α and TR3 mutants. The DBD, LBD, and A-F domains are indicated. (B/C) GST-pull-down assays for determination of RXR α /TR3 heterodimerization. GST-TR3, GST-RXR α or GST control protein immobilized on glutathione-Sepharose (20 l) was incubated with *in vitro* synthesized ³⁵S-labeled TR3 or RXR α (10 l) as indicated. Bound proteins were analyzed by SDS-PAGE/autoradiography. (D) Co-IP assay for TR3 and RXR α /385 interaction. Expression vectors for Flag epitope tagged-TR3 (Flag-TR3) and GFP-RXR α /385 were cotransfected into HEK293T cells. The expressed Flag-TR3 and GFP-RXR α /385 were then immunoprecipitated by using either anti-Flag antibody or control IgG, and immunoprecipitates were examined by western blotting using anti-GFP antibody. The same membranes were also blotted with anti-Flag antibody to determine precipitation specificity and efficiency. Input represents 5% of the total cell extract used in the precipitation assays. (E) Reporter gene assay. (NurRE)₂-tk-CAT (100 ng), β -galactosidase (100 ng) and TR3 (25 ng) expression vectors were transiently transfected into HEK293T cells together with or without RXR α or RXR α /385. Cells were treated with or without 9-*cis*-RA (10⁻⁷ M) as indicated. CAT activity was determined and

To determine how RXR ligands influence RXR α /TR3 heterodimer binding to the β RARE, RXR α protein was exposed to its ligand SR11237 or 9-*cis*-RA prior to incubation with TR3. Although the RXR α /TR3 heterodimer bound to the β RARE in the absence of an RXR ligand, the addition of SR11237 or 9-*cis*-RA strongly enhanced binding by the heterodimer (Figure 6C). These data suggest that RXR α /TR3 heterodimers in solution may be incompetent for DNA binding and that RXR ligand binding may induce a C-terminal-stabilized heterodimer conformation that favors DNA binding. In GST pull-down assays, incubation of GST-RXR α with 9-*cis*-RA strongly reduced the ability of GST-RXR α to interact with TR3/467 (Figure 6D), suggesting that 9-*cis*-RA binding may mask or alter the DBD dimerization interface required for RXR α binding to TR3 in solution. In comparison, the addition of 9-*cis*-RA did not affect the binding of RXR α /385 to GST-TR3. Thus, the dimerization interface for the formation of the RXR α /TR3 heterodimer in solution is different from that required for the formation of the DNA-bound heterodimer and is regulated by RXR ligand binding (Fig. 11).



10

glutathione-Sepharose (20 l) were incubated with *in vitro* synthesized ^{35}S -labeled receptor protein (10 l). Bound proteins were analyzed by SDS-PAGE/autoradiography.

RXR α mitochondrial localization is inhibited by RXR α ligands. Inhibition of RXR α mitochondrial localization by its ligands was demonstrated by immunoblotting of mitochondria-enriched heavy membrane (HM) fractions from LNCaP cells. The accumulation of RXR α in the HM fraction after cells were treated with TPA or an analog of AHPN/CD437 (SR11453) was inhibited by pretreatment with 9-*cis*-RA (Fig. 7A). Similar to its effect on RXR α , mitochondrial accumulation of TR3 was also abolished by 9-*cis*-RA pretreatment (Figure 7A). Consistently, cytochrome *c* (cyt *c*) release (Figure 7B) and apoptosis (Figure 7C) were inhibited by RXR ligands in LNCaP cells.

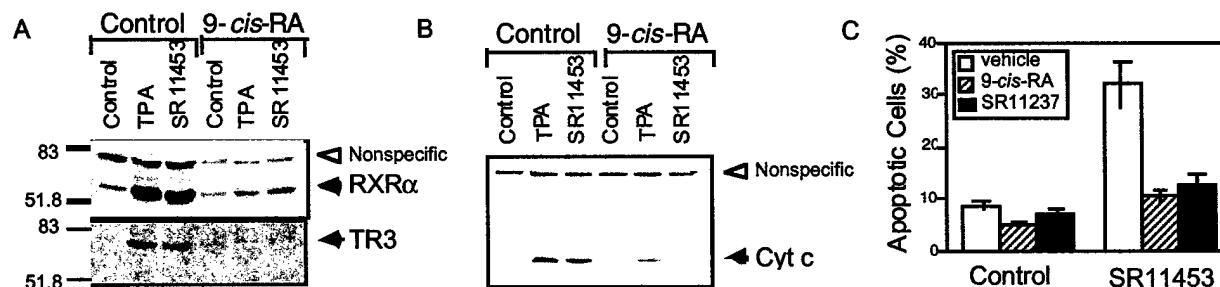


Figure 7. Effect of RXR ligands on RXR mitochondrial targeting and apoptosis. (A) Effect of 9-*cis*-RA on mitochondrial localization of RXR α and TR3. HM fractions were prepared from LNCaP cells treated with or without TPA or SR11453 (10⁻⁶ M) for 3 hr with or without a 9-*cis*-RA (10⁻⁷ M) pretreatment for 12 hr, and analyzed for expression of TR3 or RXR α by immunoblotting. (B) Inhibition of cyt *c* release by 9-*cis*-RA. LNCaP cells were treated with or without 9-*cis*-RA (10⁻⁷ M) for 12 hr before treatment with TPA (100 ng/ml) or SR11453 for 1 hr. Cytosolic fractions were analyzed for Cyt *c* by immunoblotting. A nonspecific band at ~70 kd served as a control for equal loading of proteins. (C) RXR ligands prevent SR11453 induced apoptosis. LNCaP cells were pretreated with 9-*cis*-RA or SR11237 for 12 hr before treatment with SR11453 for 48 hr. Apoptosis was determined by nuclear staining with DAPI.

2. Mitochondrial receptor of TR3

Interaction of TR3 with Bcl-2. We investigated the possibility that TR3 targeted mitochondria by binding to Bcl-2 that is known to reside on the mitochondrial outer membrane (13). GST pull-down assays showed that ^{35}S -labeled Bcl-2 was pulled-down by GST-TR3 but not by GST. Conversely, ^{35}S -labeled TR3 bound specifically to GST-Bcl-2 but not to the control GST protein (Figure 8A). We also conducted the mammalian two-hybrid assay to assess the interaction. The trans-membrane domain (TM) was deleted from Bcl-2 in order to prevent its membrane accumulation. The mutant (Bcl-2/ Δ TM) fused with the Gal4-DNA-binding domain strongly activated a reporter containing the Gal4-binding site when it was cotransfected with Gal4 transactivation-domain fused with either TR3 or TR3/ Δ DBD, a TR3 mutant lacking its DNA-binding domain (DBD) (Figure 8B). The interaction between TR3 and Bcl-2 was also demonstrated by the reporter gene assay, showing that transactivation of TR3 on its DNA responsive element was potentially inhibited by cotransfection of Bcl-2 but not by Bax (Figure 8C). To determine whether TR3 and Bcl-2 colocalized in cells, we treated cells with TPA, which induces expression of endogenous TR3 and its mitochondrial localization in LNCaP cells. Confocal microscopy analysis demonstrated that, while a low level of TR3 was detected in the nucleus of LNCaP cells in the absence of TPA, the distribution patterns of TPA-induced TR3 and Bcl-2 overlapped extensively in the cytoplasm (Figure 8D). We also determined whether transfected TR3 and Bcl-2 colocalized in cells. TR3/ Δ DBD, which constitutively resides on mitochondria in LNCaP cells, was fused to GFP. Confocal microscopy analysis demonstrated that

GFP-TR3/ Δ DBD displayed a distribution pattern that overlapped with that of transfected Bcl-2, while the control GFP protein was diffusely distributed in cells (Figure 8E). To further confirm the interaction between TR3 and Bcl-2, we generated a monoclonal antibody against the ligand-binding domain (LBD) of TR3 for use in the co-immunoprecipitation (Co-IP) assay. LNCaP cells were treated with TPA to induce TR3 expression. Cell extracts were prepared from TPA-treated and nontreated cells and incubated with anti-TR3 antibody. As shown in Figure 8F, Bcl-2 was specifically coprecipitated by anti-TR3 antibody in TPA-treated cells, but not in nontreated cells. Together, these data demonstrate that TR3 interacts with Bcl-2.

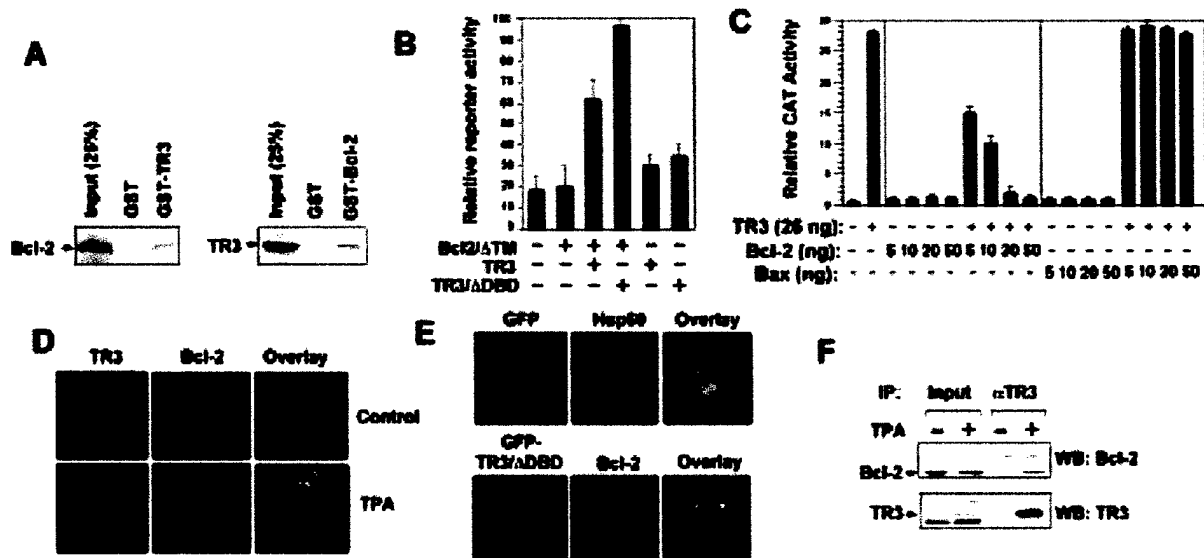


Figure 8. Interaction of TR3 with Bcl-2. (A) GST-pull down. GST-Bcl-2, GST-TR3 or GST control protein immobilized on 20 μ l of glutathione-Sepharose was incubated with 10 μ l of *in vitro* synthesized 35 S-labeled TR3 or Bcl-2 as indicated. Bound proteins were analyzed by SDS-PAGE autoradiography. (B) Mammalian two-hybrid assay. The trans-membrane (TM) domain was deleted from Bcl-2 (Bcl-2/ Δ TM) to prevent its membrane accumulation for these experiments. Gal-4 reporter gene (Gal-4-*tk*-Luc, 250 ng) and 50 ng of β -galactosidase expression vector were co-transfected into CV-1 cells with Bcl-2/ Δ TM fused with the Gal-4 DNA-binding domain (DBD) together with or without TR3 or TR3/ Δ DBD fused with the Gal-4 transactivation domain. Reporter gene activity was determined 24 h after transfection and was normalized relative to β -galactosidase activity. (C) Inhibition of TR3-dependent transactivation by Bcl-2. CV-1 cells were transfected with the NurRE-*tk*-CAT (100 ng) and β -galactosidase expression vector (100 ng) with or without TR3 expression vector (25 ng) together with or without the indicated amount of Bcl-2 or Bax expression vector. CAT activity was then determined 24 h after transfection and normalized relative to β -galactosidase activity. The bars in B and C are means \pm S.D. from three and six experiments, respectively. (D) Confocal microscopy analysis for co-localization of endogenous TR3 and Bcl-2. LNCaP cells were treated with or without TPA for 3 h, then immunostained with anti-Bcl-2 antibody followed by FITC-conjugated secondary antibody, or with mouse monoclonal anti-TR3 antibody followed by Cy3-conjugated secondary antibody. Bcl-2 and TR3 were visualized using confocal microscopy and the images were overlaid (Overlay) as indicated. (E) Confocal microscopy analysis for colocalization of transfected GFP-TR3/ Δ DBD and Bcl-2. Expression vectors for GFP-TR3/ Δ DBD and Bcl-2 were cotransfected into LNCaP cells. After 20 h, cells were immunostained with anti-Bcl-2 antibody followed by Cy3-conjugated secondary antibodies (Sigma). GFP-fusion and Bcl-2 were visualized using confocal microscopy, and the two images were overlaid. As a control, the cellular distribution of transfected GFP empty vector is shown. (F) *In vivo* co-immunoprecipitation assay. LNCaP cells were treated with or without TPA for 3 hr to induce TR3 expression, and cell lysates were incubated with mouse

monoclonal anti-TR3 antibody. After immunoprecipitation, Western blotting was conducted by using anti-Bcl-2 antibody (Santa Cruz) or rabbit polyclonal anti-TR3 anti-body (Active Motif).

Bcl-2 mediates TR3 mitochondrial targeting and apoptosis. We studied whether mutant TR3/ Δ DBD lacking the DNA-binding domain targets mitochondria by its interaction with Bcl-2. TR3/ Δ DBD expressed in HEK293T cells had a diffuse distribution pattern (Figure 9A), but colocalized with Bcl-2 and heat shock protein 60 (Hsp60), a mitochondrial specific protein, when Bcl-2 and TR3/ Δ DBD were coexpressed (Figure 9A). Immunoblotting of the mitochondria-enriched HM fractions showed a significantly enhanced accumulation of TR3/ Δ DBD in mitochondria when Bcl-2 was coexpressed (Figure 9B). These results demonstrate that Bcl-2 acts as a receptor for TR3 to target mitochondria. We next studied the involvement of TR3/Bcl-2 interaction in TR3-induced cytochrome (cyt) *c* release. In the absence of Bcl-2 cotransfection, TR3/ Δ DBD did not release cyt *c* from mitochondria as determined by confocal microscopy analysis (Figure 9C). However, after Bcl-2 cotransfection, TR3/ Δ DBD and Bcl-2 colocalized and cyt *c* was released (Figure 9C). DAPI staining indicated that TR3/ Δ DBD only caused the apoptotic events of nuclear fragmentation and condensation when Bcl-2 was cotransfected (Figure 9D). Thus, TR3/ Δ DBD-Bcl-2 interaction is required to induce cyt *c* release and apoptosis.

Our findings that Bcl-2 acts as a bridging factor allowing TR3 mitochondrial localization and apoptosis indicate that Bcl-2 does not act as an antiapoptotic protein in TR3-mediated apoptosis, but rather as a proapoptotic molecule. Our mutational analysis (data not shown) suggests that the TR3/Bcl-2 interaction converts Bcl-2 from an antiapoptotic to a proapoptotic molecule by inducing a Bcl-2 conformational change. This exciting finding further demonstrates that TR3 represents an ideal molecular target for developing cancer preventive and therapeutic drugs, since Bcl-2 is often overexpressed in cancer cells, including prostate cancer cells and is associated with multidrug resistance. In fact, overexpression of TR3 is associated with the favorable response of patients with diffuse large B-cell lymphoma to several chemotherapeutic agents (14).

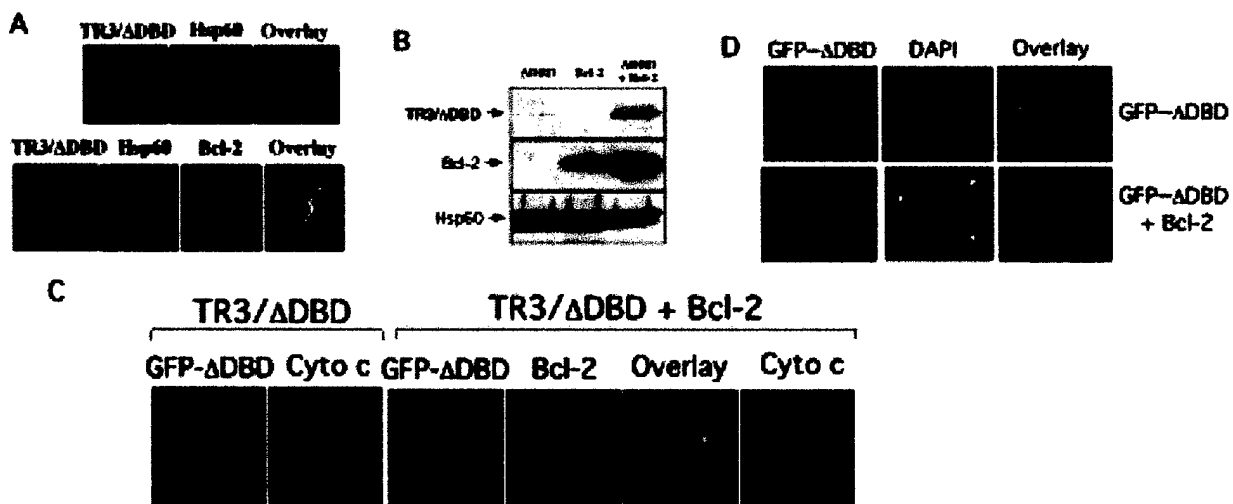


Figure 9. Bcl-2 expression promotes TR3/ Δ DBD mitochondrial localization and its apoptotic effect. (A). Confocal microscopy analysis. GFP-TR3/ Δ DBD and Bcl-2 were transfected into HEK293T cells alone or together. Cells were immunostained with anti-Bcl-2 antibody followed by Cy3-conjugated secondary antibody, or with anti-Hsp60 antibody followed by Cy5-conjugated

secondary antibody. Bcl-2, TR3/ Δ DBD and mitochondria (Hsp60) were visualized using confocal microscopy. (B). Western blotting. The heavy membrane (HM) fractions were prepared and analyzed for TR3/ Δ DBD by Western blotting using anti-GFP antibody. (C). Bcl-2 is required for TR3 to induce cyto *c* release. GFP-TR3/ Δ DBD and Bcl-2 were transfected into 293T cells alone or together. Cells were immunostained with anti-cyto *c* (cyt *c*) antibody followed by Cy5-conjugated secondary antibody, or with anti-Hsp60 followed by Cy3-conjugated secondary antibody. Cyto *c*, TR3/ Δ DBD, and mitochondria (Hsp60) were visualized using confocal microscopy. (D). Bcl-2 is required for TR3/ Δ DBD to induce apoptosis. Nuclei of cells transfected with GFP-TR3/ Δ DBD alone or with Bcl-2 were stained by DAPI. GFP-TR3/ Δ DBD expression and nuclear morphology were visualized by fluorescence microscopy. Arrows indicate cells displaying nuclear condensation and fragmentation.

3. Regulation of RAR β expression by RXR ligands.

We previously reported that RXR ligands could effectively induce RAR β expression in breast cancer cells (12). In studying how RXR ligands induce RAR β expression, we have reported that TR3/RXR heterodimer could bind effectively to the β RARE in the RAR β promoter and induced RAR β expression in response to RXR ligands. Interestingly, we also observed that RXR could bind to the β RARE as an RXR/PPAR γ heterodimer (Figure 10A). Moreover, Western analysis showed that the RXR and PPAR γ ligands were capable of inducing RAR β protein expression in ZR-75-1 and T-47D breast cancer cells (Figure 10B). Together, these data demonstrate that RXR ligands can induce RAR β expression through its activation of either RXR/TR3 or RXR/PPAR γ heterodimer.

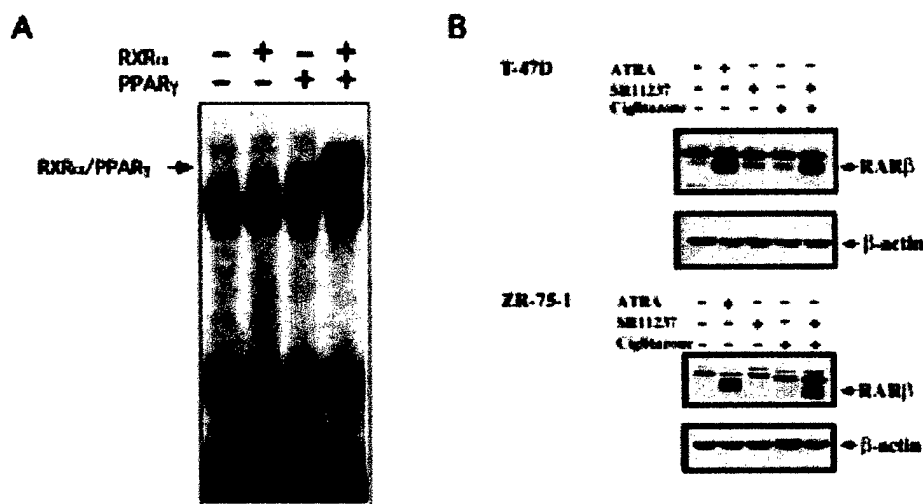


Figure 10. Regulation of RAR β expression by RXR and its ligands. A. RXR and PPAR γ bind to β RARE as a heterodimeric complex. *In vitro* translated RXR and PPAR γ were incubated with 32 P-radiolabelled β RARE either alone or in combination. The resulting reactions were then analyzed by EMSA. **B.** Regulation of RAR β protein expression by RXR and PPAR γ ligands in T-47D and ZR-75-1 breast cancer cells. T-47D and ZR-75-1 cells were treated for 24 h with SR11237 (1 μ M) in the absence or presence of ciglitazone (10 μ M). Cell lysates were prepared and Western analysis performed as described in Materials and Methods.

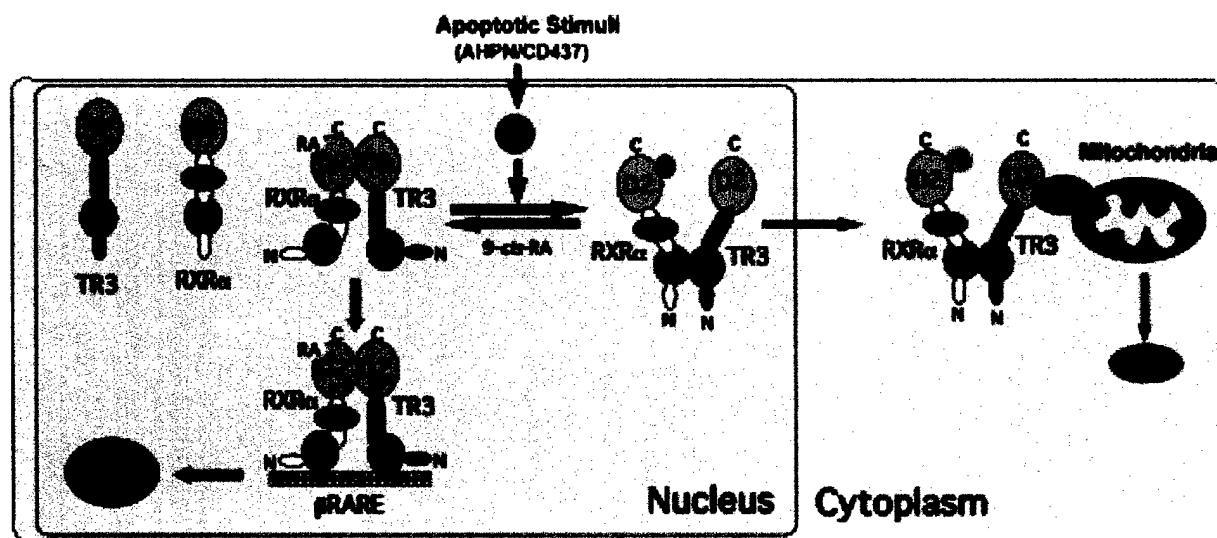


Figure 11. Mode of TR3 action supported by results obtained. Our results demonstrate that migration of TR3 from the nucleus to the cytoplasm requires its heterodimerization with RXR α . In response to an apoptotic stimulus, such as the retinoid AHPN, RXR or TR3 is modified probably through phosphorylation and then heterodimerize through their DNA-binding domain dimerization interfaces (D1), leading to the translocation of the RXR/TR3 heterodimer from the nucleus to the cytoplasm, where the heterodimer targets mitochondria through its interaction with Bcl-2. Mitochondrial targeting of the heterodimer then modulates mitochondrial function, resulting in the release of cytochrome c (cyt c) from the mitochondria into cytosol. In contrast, binding of RXR ligand 9-*cis*-RA to the RXR/TR3 heterodimer induces an RXR conformation that allows heterodimerization with TR3 through their ligand-binding domain interfaces (D2). Such a heterodimer favors DNA binding and may result in induction of RAR β , a potent growth inhibitor, through its binding to the β RARE in the RAR β promoter.

REPORTABLE OUTCOMES

1. Lin, F., Kolluri, S., Chen, G.-q. and Zhang, X.-k. Regulation of Retinoic Acid-induced Inhibition of AP-1 Activity by Orphan Receptor COUP-TF. *J. Biol. Chem* 277: 21414-22, 2002.
2. Zhang, Y., Mohammad, R., Rishi, A.K., Farhana, L., Dawson, M.I., Feng, K.-C., Leid, M., Perterson, V., Zhang, X.-k., Edelstein, M., Eilander, D., Biggar, S., Wall, N., Reichert, U., and Fontana, J.A. Induction of apoptosis of human B-CLL and ALL cells by a novel retinoid and its non-retinoidal analog. *Blood*. 100: 2917-2925, 2002.
3. Dawson MI, Zhang X.-k. Discovery and design of retinoic acid receptor and retinoid X receptor class- and subtype-selective synthetic analogs of all-*trans*-retinoic acid and 9-*cis*-retinoic acid. *Curr. Med. Chem.* 9: 623-37. 2002.
4. James, S., Lin, F., Kolluri, S., Dawson, M.I., and Zhang, X.-k. Regulation of retinoic acid receptor b expression by peroxisome proliferator-activated receptor γ ligands in cancer cells. *Cancer Research* 63, 3531-3538, 2003.

CONCLUSIONS

We have investigated the mechanism by which retinoids induce apoptosis in breast cancer cells. Our results demonstrate that retinoid AHPN and its analogs exert their apoptotic effect by inducing migration of orphan receptor TR3 from the nucleus to mitochondria. In studying how the migration is regulated, we have observed that the migration of TR3 from the nucleus to the cytoplasm requires RXR α through their heterodimerization. We have also found that RXR contains a nuclear export sequence that is required for its cytoplasmic localization and is regulated by RXR ligands. Moreover, we have demonstrated a unique TR3/RXR α heterodimerization, which allows efficient nuclear export of the heterodimer. These data not only enhance our understanding the molecular mechanism by which TR3 translocates from the nucleus to the cytoplasm but also provide a molecular basis for developing RXR ligands that regulate apoptosis of breast cancer cells. In studying how TR3 targets mitochondria, we have discovered that TR3 can physically interacts with Bcl-2 and that Bcl-2 acts as a mitochondrial receptor of TR3 and is required for apoptotic effect of TR3 in breast cancer cells. Our finding that Bcl-2 is apoptotic when it interacts with TR3 provides novel approach to induce apoptosis of Bcl-2-expressing breast cancer cells. The overall plan for the next year of research remains the same as proposed in the original grant application.

REFERENCES

1. Shao, Z. M., Dawson, M. I., Li, X. S., Rishi, A. K., Sheikh, M. S., Han, Q. X., Ordonez, J. V., Shroot, B., and Fontana, J. A. p53 independent G0/G1 arrest and apoptosis induced by a novel retinoid in human breast cancer cells. *Oncogene*, 11: 493-504, 1995.
2. Li, Y., Lin, B., Agadir, A., Liu, R., Dawson, M. I., Reed, J. C., Fontana, J. A., Bost, F., Hobbs, P. D., Zheng, Y., Chen, G. Q., Shroot, B., Mercola, D., and Zhang, X. K. Molecular determinants of AHPN (CD437)-induced growth arrest and apoptosis in human lung cancer cell lines. *Mol Cell Biol*, 18: 4719-4731, 1998.
3. Li, H., Kolluri, S. K., Gu, J., Dawson, M. I., Cao, X., Hobbs, P. D., Lin, B., Chen, G., Lu, J., Lin, F., Xie, Z., Fontana, J. A., Reed, J. C., and Zhang, X. Cytochrome c release and apoptosis induced by mitochondrial targeting of nuclear orphan receptor TR3 [see comments] [comment]. *Science*, 289: 1159-1164, 2000.
4. Dawson, M. I., Hobbs, P. D., Peterson, V. J., Leid, M., Lange, C. W., Feng, K. C., Chen, G., Gu, J., Li, H., Kolluri, S. K., Zhang, X., Zhang, Y., and Fontana, J. A. Apoptosis induction in cancer cells by a novel analogue of 6-[3-(1- adamantyl)-4-hydroxyphenyl]-2-naphthalenecarboxylic acid lacking retinoid receptor transcriptional activation activity. *Cancer Res*, 61: 4723-4730., 2001.
5. Forman, B. M., Umesono, K., Chen, J., and Evans, R. M. Unique response pathways are established by allosteric interactions among nuclear hormone receptors. *Cell*, 81: 541-550, 1995.
6. Perlmann, T. and Jansson, L. A novel pathway for vitamin A signaling mediated by RXR heterodimerization with NGFI-B and NURR1. *Genes Dev*, 9: 769-782, 1995.
7. Zhang, Y., Dawson, M. I., Mohammad, R., Rishi, A. K., Farhana, L., Feng, K. C., Leid, M., Peterson, V., Zhang, X. K., Edelstein, M., Eilander, D., Biggar, S., Wall, N., Reichert, U., and Fontana, J. A. Induction of apoptosis of human B-CLL and ALL cells by a novel retinoid and its nonretinoidal analog. *Blood*, 100: 2917-2925, 2002.
8. Zhang, X. K., Salbert, G., Lee, M. O., and Pfahl, M. Mutations that alter ligand-induced switches and dimerization activities in the retinoid X receptor. *Mol Cell Biol*, 14: 4311-4323, 1994.
9. Bourguet, W., Ruff, M., Chambon, P., Gronemeyer, H., and Moras, D. Crystal structure of the ligand-binding domain of the human nuclear receptor RXR- α . *Nature*, 375: 377-382, 1995.
10. Bourguet, W., Vivat, V., Wurtz, J. M., Chambon, P., Gronemeyer, H., and Moras, D. Crystal structure of a heterodimeric complex of RAR and RXR ligand- binding domains. *Mol Cell*, 5: 289-298., 2000.

11. Meinke, G. and Sigler, P. B. DNA-binding mechanism of the monomeric orphan nuclear receptor NGFI-B. *Nat Struct Biol*, 6: 471-477, 1999.
12. Wu, Q., Dawson, M. I., Zheng, Y., Hobbs, P. D., Agadir, A., Jong, L., Li, Y., Liu, R., Lin, B., and Zhang, X. K. Inhibition of trans-retinoic acid-resistant human breast cancer cell growth by retinoid X receptor-selective retinoids. *Mol Cell Biol*, 17: 6598-6608, 1997.
13. Reed, J. C. Bcl-2 family proteins. *Oncogene*, 17: 3225-3236, 1998.
14. Shipp, M. A., Ross, K. N., Tamayo, P., Weng, A. P., Kutok, J. L., Aguiar, R. C., Gaasenbeek, M., Angelo, M., Reich, M., Pinkus, G. S., Ray, T. S., Koval, M. A., Last, K. W., Norton, A., Lister, T. A., Mesirov, J., Neubergh, D. S., Lander, E. S., Aster, J. C., and Golub, T. R. Diffuse large B-cell lymphoma outcome prediction by gene-expression profiling and supervised machine learning. *Nat Med*, 8: 68-74., 2002.

APPENDICES

Lin, F., Kolluri, S., Chen, G.-q. and Zhang, X.-k. Regulation of Retinoic Acid-induced Inhibition of AP-1 Activity by Orphan Receptor COUP-TF. *J. Biol. Chem* 277: 21414-22, 2002.

James, S., Lin, F., Kolluri, S., Dawson, M.I., and Zhang, X.-k. Regulation of retinoic acid receptor β expression by peroxisome proliferator-activated receptor γ ligands in cancer cells. *Cancer Research* 63, 3531-3538, 2003.

Regulation of Retinoic Acid-induced Inhibition of AP-1 Activity by Orphan Receptor Chicken Ovalbumin Upstream Promoter-Transcription Factor*

Received for publication, February 25, 2002, and in revised form, April 2, 2002
Published, JBC Papers in Press, April 4, 2002, DOI 10.1074/jbc.M201885200

Feng Lin, Siva Kumar Kolluri, Guo-quan Chen, and Xiao-kun Zhang†

From the Burnham Institute, Cancer Center, La Jolla, California 92037

Retinoids are therapeutically effective in the treatment of various cancers, and some of the therapeutic action of retinoids can be ascribed to their potent inhibition of AP-1 activity that regulates transcription of genes associated with cell growth. We recently reported that the expression of orphan receptor chicken ovalbumin upstream promoter-transcription factor (COUP-TF) plays a role in mediating the growth inhibitory effect of *trans*-retinoic acid (*trans*-RA) in cancer cells. To gain insight into the molecular mechanism by which COUP-TF regulates *trans*-RA activity, we evaluated the effect of COUP-TF on antagonism of AP-1 activity by *trans*-RA. Our results demonstrated a positive correlation between COUP-TF expression and the ability of *trans*-RA to inhibit AP-1 activity in various cancer cell lines. In transient transfection assay, expression of COUP-TF strongly inhibited tumor promoter 12-*O*-tetradecanoylphorbol-13-acetate-induced AP-1 transactivation activity and transactivation of c-Jun/c-Fos in both a *trans*-RA-dependent and -independent manner. *In vitro* studies demonstrated that the addition of COUP-TF inhibited c-Jun DNA binding through a direct protein-protein interaction that is mediated by the DNA binding domain of COUP-TF and the leucine zipper of c-Jun. Stable expression of COUP-TF in COUP-TF-negative MDA-MB231 breast cancer cells restored the ability of *trans*-RA to inhibit 12-*O*-tetradecanoylphorbol-13-acetate-induced c-Jun expression. The effect of COUP-TF in enhancing the *trans*-RA-induced antagonism of AP-1 activity required expression of retinoic acid receptors (RARs), since stable expression of COUP-TF in COUP-TF-negative HT-1376 bladder cancer cells, which do not express RAR α and RAR β , failed to restore *trans*-RA-induced AP-1 repression. Thus, COUP-TF, through its physical interaction with AP-1, promotes anticancer effects of retinoids by potentiating their anti-AP-1 activity.

nuclear receptor classes, the retinoic acid receptor (RAR)¹ and retinoid X receptor (RXR) (3–5). Both are encoded by three different genes (α , β , and γ) and function as ligand-inducible transcription factors *in vivo* mainly as RXR/RAR heterodimers. *trans*-Retinoic acid (*trans*-RA) binds RARs, whereas 9-*cis*-RA binds both RARs and RXRs. Binding of retinoids to their receptors induces receptor conformational changes that switch on transcription of genes containing RA response elements (RAREs) (3–5). In addition to their positive regulation of RARE-containing genes, retinoid receptors, in response to their ligands, can inhibit effects induced by the tumor promoter 12-*O*-tetradecanoylphorbol-13-acetate (TPA) and the transcriptional activity of the proto-oncogenes c-Jun and c-Fos (6), which are components of the AP-1 complex, which often has a role in cancer cell proliferation (7). The activation of AP-1-responsive genes by TPA or c-Jun/c-Fos through TPA response elements (TREs) is repressed by retinoid receptors in a ligand-dependent manner (8–10). Conversely, AP-1 represses transactivation of retinoid receptors (8, 9). This mutual antagonism appears to play a critical role in regulating cell growth and differentiation (6). For example, overexpression of c-Jun conferred retinoid resistance to breast cancer cells (11), while overexpression of retinoid receptors enabled *trans*-RA to inhibit AP-1 activity in ovarian cancer cells and their growth (12). The functional interaction between AP-1 and retinoid receptors is also observed for other nuclear receptors, including glucocorticoid receptor (13–16), thyroid hormone receptor (17), vitamin D receptor, estrogen receptor (18, 19), and androgen receptor (AR) (18).

The anti-AP-1 activities shown by liganded retinoid receptors appear to contribute significantly to the therapeutic efficacy of retinoids against hyperproliferative diseases (6). Transcription of various AP-1-responsive genes, such as collagenase and stromelysin, which have roles in tumor progression and invasiveness (20), is inhibited by retinoids (8–10) and has been reported to contribute to their reversal of human bronchial epithelial squamous differentiation (21). Retinyl methyl ether, which effectively prevents mammary cancer in animals, strongly suppressed AP-1 activity in breast cancer cell (22). Interestingly, RAR β , a negative regulator of cancer cell growth (23), potently inhibits AP-1 activity and collagenase expression in both breast and lung cancer cells (24). Recent studies demonstrate that retinoids that specifically inhibit AP-1 activity but antagonize RAR transactivation on RAREs inhibited the growth of many different types of cancer cells (25–27). Anti-AP-1 retinoids inhibited squamous differentiation of human

Retinoids, the natural and synthetic vitamin A analogs, exert profound effects on many biological processes, including cell proliferation and differentiation (1, 2), and are recognized as promising agents for the prevention and treatment of various cancers. The effects of retinoids are mainly mediated by two

* This work is supported in part by grants from the National Institutes of Health and the United States Army Medical Research and Materiel Command (to X. Z.). The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

† To whom correspondence should be addressed: Burnham Institute, Cancer Center, 10901 North Torrey Pines Rd., La Jolla, CA 92037. Tel.: 858-646-3141; Fax: 858-646-3195; E-mail: xzhang@burnham.org.

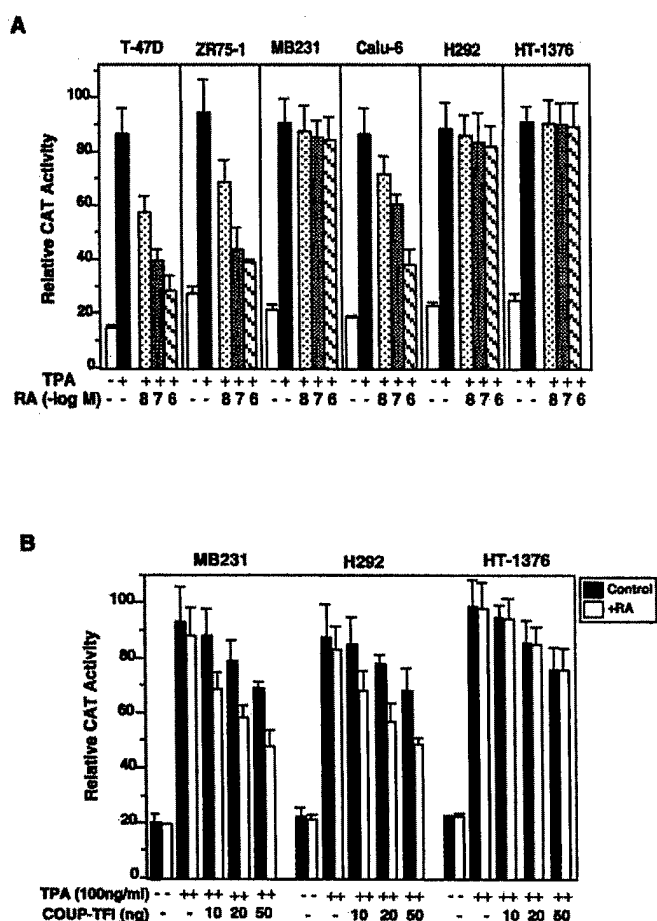


FIG. 1. COUP-TF expression and *trans*-RA-induced anti-AP-1 activity correlate in cancer cell lines. A, inhibition of TPA-induced collagenase promoter activity by *trans*-RA. The -73Col-CAT reporter was transfected into the indicated cancer cell lines. After transfection, the cells were incubated in DMEM medium containing 0.5% FCS for 24 h and treated with either TPA (100 ng/ml) alone or with the indicated concentrations of *trans*-RA. After 12 h, the cells were harvested, and CAT activity was determined. The activities of cotransfected β -galactosidase were used as controls for transfection efficiency. B, effect of COUP-TF expression in COUP-TF-negative cancer cell lines on inhibition of AP-1 activity by *trans*-RA. The -73Col-CAT reporter was transfected with or without COUP-TF into COUP-TF-negative cancer cell lines (MDA-MB231, H292, and HT-1376). Cells were incubated in medium (see "Experimental Procedures") containing 0.5% FCS for 24 h and then treated with either TPA (100 ng/ml) alone or with 10^{-6} M *trans*-RA. After 12 h, cells were harvested, and CAT activity was determined. The activities of cotransfected β -galactosidase were used as reference values.

bronchial epithelial cells (21), TPA-induced transformation and the clonal growth of the promotion-sensitive JB6 mouse epidermal cell line (28), and papilloma formation in animals (29). Thus, anti-AP-1 activity of retinoids contributes to their chemopreventive and chemotherapeutic effects, presumably by blocking the processes of tumor promotion and cell transformation.

The mechanism by which retinoids inhibit AP-1 activity remains largely unclear. Unlike the effect of retinoid receptors on RAREs, inhibition of AP-1 activity by retinoid receptors is independent of retinoid receptor-RARE interaction (8–10). Previous studies suggested several possible mechanisms for AP-1 inhibition by retinoid receptors. First, retinoid receptors are reported to physically interact with c-Jun and/or c-Fos (8, 9). This interaction results in the mutual inhibition of their DNA binding and transactivation functions and could explain the cross-talk occurring between AP-1 and retinoid signaling. However, a large excess of either retinoid receptor protein or c-Jun

and c-Fos proteins was required to inhibit *in vitro* binding to the TRE or RARE, respectively (8, 9). Because the *in vivo* footprint assay for glucocorticoid receptor and AP-1 interaction did not reveal any effect on DNA binding (30), whether RAR and AP-1 directly interact *in vivo* remains to be established. Subsequently, it was suggested that blocking activation of the Jun N-terminal kinase signaling pathway that activates AP-1 might be responsible for AP-1 inhibition by retinoid and other nuclear receptors (31). Although this mechanism may explain how retinoids inhibit AP-1 activity under some conditions, it does not address how mutual inhibition occurs. Recent results suggest that the molecular basis of receptor-mediated inhibition of AP-1 transcriptional activation might be due to competition for a common coactivator, such as the cAMP-response element-binding protein (CBP), which is required for transcriptional activation by both the AP-1 complex and RARs (32). However, a domain of RAR capable of inhibiting AP-1 activity, such as the DNA-binding domain (33), does not interact with CBP (32). The development of retinoids that specifically inhibit AP-1 activity without transactivating RARs (25, 26) also argues against the involvement of RAR coactivators in RARs-AP-1 cross-talk. Thus, the availability of CBP is unlikely to be the sole modulator of RAR and AP-1-signaling pathways, and other adapter proteins may be involved in the antagonism of AP-1 activity by RARs.

Recent studies demonstrate that orphan receptor COUP-TF is involved in regulation retinoid responses (34, 35). COUP-TF is encoded by two distinct genes, COUP-TFI (EAR-3) (36, 37) and COUP-TFII (ARP-1) (38). Both show exceptional homology and overlapping expression patterns, suggesting their redundant functions (39). COUP-TF can modulate retinoid responses through either its high affinity binding to various RAREs or its heterodimerization with RXR (40–43). We previously reported that COUP-TF expression was required for cancer cell growth inhibition by *trans*-RA (35). We further demonstrated that the effect of COUP-TF is partly due to its induction of RAR β that mediates growth inhibition by retinoids in various cancer cells (35).

To further understand how COUP-TF is involved in the regulation of *trans*-RA activity in cancer cells, we investigated the effect of COUP-TF on antagonism of AP-1 activity by *trans*-RA. Our data demonstrate that COUP-TF expression in various cancer cell lines correlated with the ability of *trans*-RA to suppress AP-1 transcriptional activity. In addition, we found that COUP-TF effectively suppressed AP-1 transcriptional activity by interacting with c-Jun to cause loss of c-Jun DNA binding. This interaction required COUP-TF DNA-binding domain and the c-Jun leucine-zipper domain. Although AP-1 inhibition by COUP-TF did not require *trans*-RA, COUP-TF strongly potentiated the AP-1 antagonism by *trans*-RA when RARs were expressed. Our results demonstrate that interaction between COUP-TF and AP-1 is involved in regulating anti-AP-1 activity of retinoids in cancer cells and suggest that COUP-TF plays a role in the cross-talk between retinoid and AP-1 signalings.

EXPERIMENTAL PROCEDURES

Cell Culture—HeLa ovarian and MDA-MB231 breast cancer cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal calf serum (FCS); Calu-6 lung cancer and HT-1376 bladder cancer cells were grown in MEM containing 10% FCS; and T-47D and ZR-75-1 breast cancer and H292 lung cancer cells were grown in RPMI-1460 medium with 10% FCS.

Plasmid Construction—CAT reporter constructs TRE-tk-CAT and -73Col-CAT have been described (8, 17, 22, 24), as have expression vectors for RAR α , COUP-TF, c-Jun, and c-Fos (8, 17, 22, 24, 44). pcDNA3-COUP-TFII C-terminal deletion mutants pcDNA3-COUP-TFII Δ 7, pcDNA3-COUP-TFII Δ 30, and pcDNA3-COUP-TFII Δ 108 were generated as described (35). pcDNA3-COUP-TFII N-terminal deletion mutants were

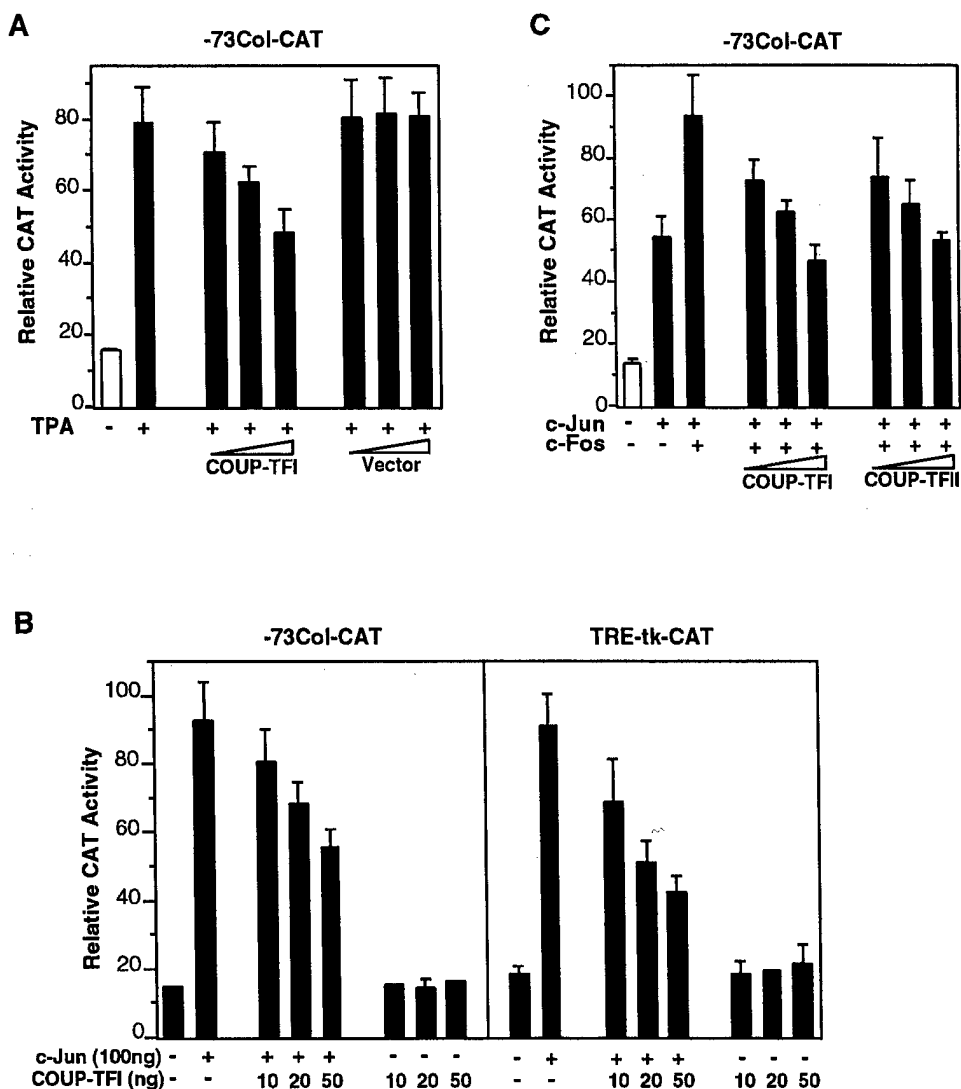


FIG. 2. Inhibition of AP-1 activity by COUP-TF. A, inhibition of TPA-induced collagenase promoter activity by COUP-TF in HeLa cells. The -73Col-CAT reporter was cotransfected without or with COUP-TFI expression vector (10, 20, or 50 ng) or the control pcDNA3 vector into cells. After transfection, cells were incubated in DMEM medium containing 0.5% FCS for 24 h and then treated with or without TPA (100 ng/ml). After 12 h, the cells were harvested, and CAT activity was determined. B, inhibition of c-Jun-induced -73Col-CAT and TRE-tk-CAT activity by COUP-TF in HeLa cells. The -73Col-CAT or TRE-tk-CAT reporter was cotransfected with/without c-Jun in the presence or absence of COUP-TFI expression vector (10, 20, and 50 ng) into cells. After transfection, cells were incubated in DMEM medium containing 0.5% FCS for 24 h, and CAT activity was determined. C, inhibition of c-Jun and c-Fos activities by COUP-TF in HeLa cells. The -73Col-CAT reporter was cotransfected with/without c-Jun (50 ng) and/or c-Fos (50 ng) expression vectors either in the presence or absence of COUP-TFI or COUP-TFII expression vector (10, 20, and 50 ng) into HeLa cells. The cells were then harvested, and CAT activity was determined.

constructed by cloning PCR products from COUP-TFII into pcDNA3 (Stratagene) using the following forward primers: TATAGGTACCATGGCGCCGCCGTGCCC for pcDNA3-COUP-TFIIΔN25; ATATGGTACCATGACGCCAGCCCAGACG for pcDNA3-COUP-TFIIΔN50; AATTGGTACCATGCACATCGAGTGCCTG for pcDNA3-COUP-TFIIΔN75; and ATTAGGTACCATGCACCATCGCAACCAG for pcDNA3-COUP-TFIIΔN125. The oligonucleotide GTGGCAGTTGAGGGGATCC was used as the reverse primer for these mutants.

Transient and Stable Transfection Assays—HeLa cells (1×10^5 cells/well) were plated in 24-well plates for 16–24 h before transfection as described (34, 35, 45), and other cancer cells (5×10^5 cell/well) were seeded in six-well plates. A modified calcium phosphate precipitation procedure was used for transient transfections (34, 35, 45). Briefly, 200 ng of reporter plasmid, 100 ng of β -galactosidase of expression vector (pCH 110; Amersham Biosciences), and various amounts of each expression vector were mixed with carrier DNA (pBluescript) to give 1000 ng of total DNA/well. CAT activity was normalized for transfection efficiency to the responding β -Gal activity. For stable transfections, the pRC/CMV-COUP-TF recombinant was transfected into MDA-MB231 and HT-1376 cells by the calcium phosphate precipitation method, and the stable clones were screened with G418 (Invitrogen) as described (35). Integration and expression of transfected cDNA were determined

by Southern blotting and Northern blotting, respectively.

Preparation of Receptor Proteins—Receptor proteins for RAR α , TR3, COUP-TF, and its mutants were synthesized by *in vitro* transcription-translation using rabbit reticulocyte lysates (Promega) as described previously (34). Amounts of translated proteins were determined by [35 S]methionine incorporation and SDS-PAGE with quantitation by incorporated radioactivity after normalization relative to methionine content.

Gel Retardation Assay—A fragment of the collagenase promoter region -73 to +63 containing one AP-1-binding site (TRE) was excised from a collagenase-CAT construct (8). In addition, the AP-1 binding sequence 5'-GATCCGGATGAGTCACCA-3' was synthesized. Two fragments were labeled with [32 P]dCTP for use as probes for protein-DNA interaction. *In vitro* translated protein was incubated with the probe in a 20- μ l reaction mixture containing 10 mM HEPES, pH 7.9, 50 mM KCl, 1 mM dithiothreitol, 2.5 mM MgCl $_2$, 10% glycerol, and 1 μ g of poly(dI-dC) at 25 $^{\circ}$ C for 15 min. DNA-protein complexes were resolved on 5% nondenaturing polyacrylamide gels, and then gels were dried and analyzed by autoradiography.

GST Pull-down Assay—To prepare glutathione S-transferase (GST)-c-Jun or GST-c-Jun mutant fusion proteins, each c-Jun DNA or c-Jun mutant fragment was cloned in frame into the expression vector

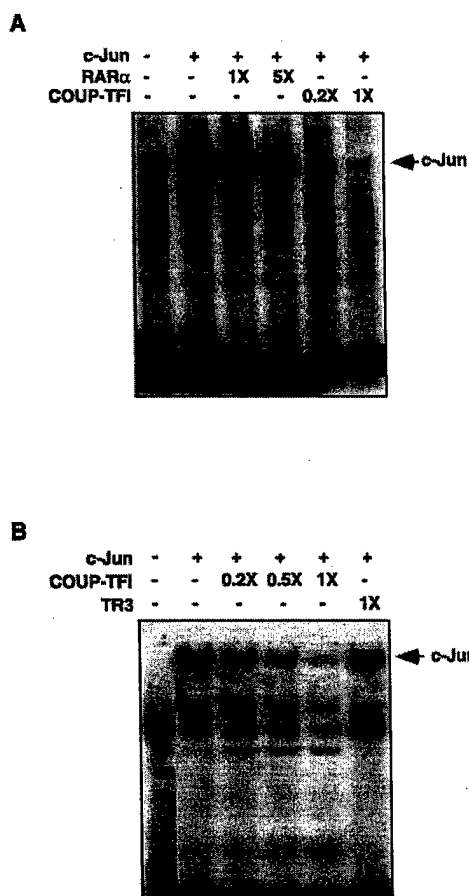


FIG. 3. Inhibition of c-Jun binding to DNA by COUP-TF. *In vitro* synthesized c-Jun was preincubated with the indicated molar excess of COUP-TF, RAR α , or TR3. Unprogrammed reticulocyte lysate was added to maintain an equal protein concentration in each reaction. Following preincubation, the reaction mixtures were incubated with 32 P-labeled collagenase promoter (A) or the TRE (B) and analyzed by gel retardation. The arrowhead indicates the c-Jun-binding complex.

pGEX-3X (Amersham Biosciences). Fusion proteins were expressed in bacteria using the manufacturer's procedure and analyzed by gel retardation assays and Western blotting (data not shown). To determine the interaction between COUP-TF and c-Jun, the fusion proteins were immobilized on glutathione-Sepharose beads. The vector protein (GST), prepared under the same conditions as a control, was also immobilized. Beads were preincubated with bovine serum albumin (1 mg/ml) at room temperature for 5 min. 35 S-labeled *in vitro* translated COUP-TF proteins (2–5 μ l, depending on translation efficiency) were then added to the beads. The beads in 200 μ l in EBC buffer (140 mM NaCl, 0.5% Nonidet P-40, 100 mM NaF, 200 mM sodium orthovanadate, and 50 mM Tris, pH 8.0) were rocked continuously for 1 h at 4 $^{\circ}$ C. After washing 5 times with NETN buffer (100 mM NaCl, 1 mM EDTA, 20 mM Tris, pH 8.0, 0.5% Nonidet P-40), bound proteins were analyzed by SDS-PAGE and autoradiography.

Northern Blotting—For Northern analysis, total RNAs were prepared using an RNeasy Mini Kit (Qiagen, Hilden, Germany). Total RNA (30 μ g) from different cell lines treated with or without *trans*-RA (10 $^{-6}$ M) in the presence or absence of TPA (100 ng/ml) was analyzed by Northern blotting as described (46).

RESULTS

Correlation between COUP-TF Expression and Inhibition of AP-1 Activity by *trans*-RA—We recently reported that expression of COUP-TF is required for growth inhibition and apoptosis induction by *trans*-RA in various cancer cells (35). Since inhibition of AP-1 activity by retinoids is known to contribute to their anticancer effects, we studied whether COUP-TF expression was involved in the process. The effect of *trans*-RA on inhibiting AP-1 activity (Fig. 1A) was evaluated in COUP-TF-

positive T-47D and ZR-75-1 breast cancer and Calu-6 lung cancer cell lines and in COUP-TF-negative MDA-MB231 breast cancer, H292 lung cancer, and HT-1376 bladder cancer cell lines (35). AP-1 activity induced by TPA was determined by transient transfection using the reporter -73Col-CAT, which contains a TRE that binds AP-1 (8). TPA-induced reporter activity was strongly inhibited in a *trans*-RA-dependent manner in the COUP-TF-positive ZR-75-1, T-47D, and Calu-6 cell lines. In contrast, *trans*-RA showed very little effect on TPA-induced -73Col-CAT activity in the COUP-TF-negative MDA-MB231, H292, and HT-1376 cell lines. Thus, COUP-TF expression correlates positively with the ability of *trans*-RA to inhibit AP-1 transcriptional activity.

Trans-RA-dependent and -independent Antagonism of AP-1 Activity by COUP-TF—The positive association between COUP-TF expression and *trans*-RA-induced anti-AP-1 activity suggested that COUP-TF was required for *trans*-RA to inhibit AP-1 activity and that the absence of COUP-TF expression in MDA-MB231, H292, and HT-1376 cells might be responsible for the lack of *trans*-RA activity. Therefore, we transiently transfected the COUP-TF expression vector and the -73Col-CAT reporter into the COUP-TF-negative cell lines, in which *trans*-RA did not inhibit AP-1 transcriptional activity (Fig. 1B). Upon COUP-TF transfection, *trans*-RA inhibited TPA-induced reporter activity in both MDA-MB231 and H292 cells in a COUP-TF concentration-dependent manner. Interestingly, COUP-TF transfection alone produced some inhibition of AP-1 activity. This observation suggested that COUP-TF inhibited AP-1 activity in both *trans*-RA-dependent and -independent manners in MDA-MB231 and H292 cells. When evaluated in HT-1376 cells, COUP-TF expression was able to inhibit AP-1 activity independently of *trans*-RA (Fig. 1B). However, it failed to confer the ability of *trans*-RA to inhibit TPA-induced reporter activity, even at high transfection levels, suggesting that *trans*-RA-dependent inhibition of AP-1 activity by COUP-TF is impaired in this cell line.

The above results suggested that COUP-TF alone inhibited AP-1 activity. We then examined the anti-AP-1 activity of COUP-TF in HeLa cells, in which TPA strongly induced the transcription of the transfected -73Col-CAT reporter. Similar to that observed in other cancer cell lines (Fig. 1B), COUP-TFI cotransfection inhibited TPA-induced reporter activity in a COUP-TFI concentration-dependent manner (Fig. 2A). Cotransfection of COUP-TFII produced almost identical results (data not shown). Activation of collagenase promoter by TPA occurs mainly through induction of AP-1 activity that activates the TRE in the promoter (7). We therefore examined whether COUP-TF expression also interfered with transactivation activity of c-Jun homodimer and c-Jun/c-Fos heterodimer. Cotransfection of -73Col-CAT with the c-Jun expression vector into HeLa cells led to about 5-fold induction of reporter expression (Fig. 2B), presumably due to activation of the collagenase promoter by the c-Jun homodimer. The c-Jun-induced -73Col-CAT reporter activity was repressed when COUP-TFI expression vector was cotransfected. Similarly, -73Col-CAT reporter activity induced by c-Jun/c-Fos heterodimers was inhibited by COUP-TFI and COUP-TFII (Fig. 2C). To determine that inhibition of collagenase promoter activity by COUP-TF was mediated by the TRE, we evaluated how COUP-TFI affected the TRE-tk-CAT reporter, which has a TRE sequence fused to the thymidine kinase promoter (Fig. 2B). Our result showed that cotransfection of COUP-TFI significantly inhibited c-Jun-induced TRE-tk-CAT activity (Fig. 2B). These results clearly demonstrate that COUP-TF inhibits transcriptional activity of c-Jun homodimer and c-Jun/c-Fos heterodimer and that the inhibition of AP-1 activity by COUP-TF on the TRE is respon-

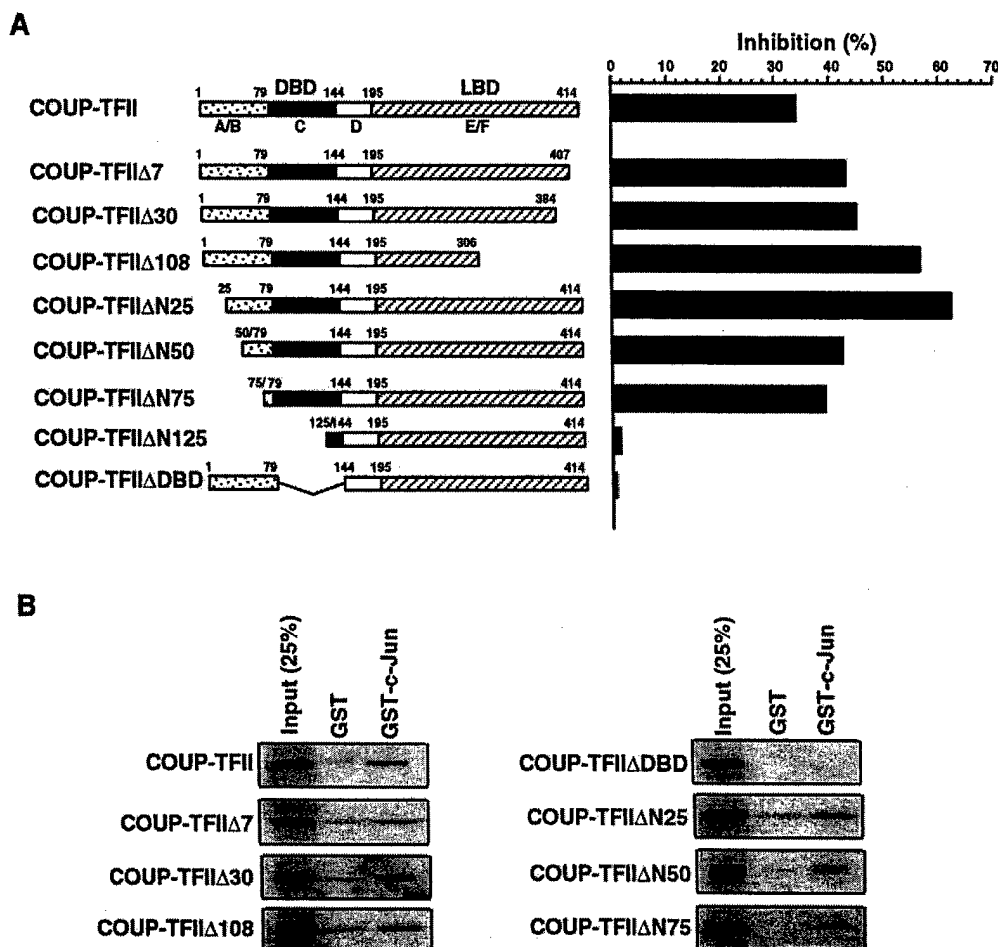


FIG. 4. Interaction of c-Jun and COUP-TF and their domain requirements. *A*, inhibition of transcriptional activity of AP-1 by COUP-TF mutants in HeLa cells. *Left*, schematic representation of COUP-TFII and its mutants. The DNA binding domain (DBD) and ligand-binding domain (LBD) of COUP-TFII are indicated. *Right*, effect of COUP-TFII mutants on AP-1 activity. The -73Col-CAT reporter was cotransfected without or with c-Jun (100 ng) alone or together with the indicated COUP-TFII mutants (50 ng). After transfection, cells were incubated in DMEM containing 0.5% FCS for 24 h. After 12 h, the cells were harvested, and CAT activity was determined. Reporter activity is shown as percentage of inhibition. *B*, interaction between c-Jun and COUP-TFs. c-Jun was synthesized in bacteria using pGEX-3X expression vector. GST-c-Jun fusion protein was immobilized on the glutathione-Sepharose beads. As a control, the same amount of glutathione *S*-transferase was also immobilized on the beads. *In vitro* translated ³⁵S-labeled COUP-TF and its mutant proteins were then mixed with the beads. After extensive washing, the bound proteins were analyzed by SDS-PAGE. The input proteins are shown for comparison.

sible for its antagonism of TPA activity.

Inhibition of c-Jun Binding to DNA by COUP-TF—Inhibition of AP-1 activity by several nuclear receptors has been shown to be due to their inhibition of AP-1 binding to DNA (8, 9, 13–15, 17, 22, 47). To study whether inhibition of AP-1 DNA binding by COUP-TF was involved in its inhibition of AP-1 activity, the purified collagenase promoter fragment was used as a probe in gel shift assays for binding of *in vitro* synthesized c-Jun protein in the absence or presence of COUP-TF protein (Fig. 3A). c-Jun alone formed a strong complex with the promoter. However, upon preincubation with COUP-TF protein, c-Jun binding was significantly inhibited. The addition of COUP-TF protein did not show any new complex formed with the collagenase promoter, indicating that the inhibitory effect of COUP-TF is not due to its direct binding to the collagenase promoter. We also studied the effect of COUP-TF on binding of c-Jun to the TRE. Preincubation with COUP-TF protein similarly inhibited c-Jun binding to the TRE derived from the collagenase promoter (Fig. 3B). As a comparison, preincubation of c-Jun with TR3 orphan receptor (34) that could not antagonize c-Jun transactivation (data not shown) did not inhibit c-Jun binding to the TRE. This result demonstrated that the inhibitory effect of COUP-TF on c-Jun binding to the collagenase promoter is not due to regions other than the TRE in the promoter. Next, we compared the

inhibitory effect of COUP-TF with that of RAR α that is known to repress AP-1 activity and inhibit c-Jun binding to TRE (8). Whereas an equal amount of COUP-TF significantly inhibited c-Jun binding to the collagenase promoter, an equal amount of RAR α did not show any detectable inhibition (Fig. 3A). Inhibition of c-Jun binding required a 5-fold excess of RAR α , similar to that observed before (8). The addition of *trans*-RA did not enhance the inhibitory effect of RAR α (Ref. 8 and data not shown). Together, these results demonstrate that the inhibition of c-Jun binding to the TRE contributes to its suppression of AP-1 transcriptional activity by COUP-TF and that COUP-TF is a more effective inhibitor than RAR α .

Interaction of c-Jun and COUP-TF—Our observation that COUP-TF inhibits transactivation and DNA binding by c-Jun prompted us to examine whether COUP-TF and c-Jun interact directly using GST pull-down assay (Fig. 4). *In vitro* synthesized radiolabeled COUP-TFII was specifically pulled down by bacterially expressed GST-c-Jun hybrid protein but not by GST protein (Fig. 4B), demonstrating that COUP-TF and c-Jun interact in solution. To identify the domain of COUP-TFII responsible for interacting with c-Jun, COUP-TFII deletion mutants (Fig. 4A) were constructed and analyzed on the -73Col-CAT reporter for antagonism of AP-1 activity in HeLa cells (Fig. 4A) and for their interaction with c-Jun using the

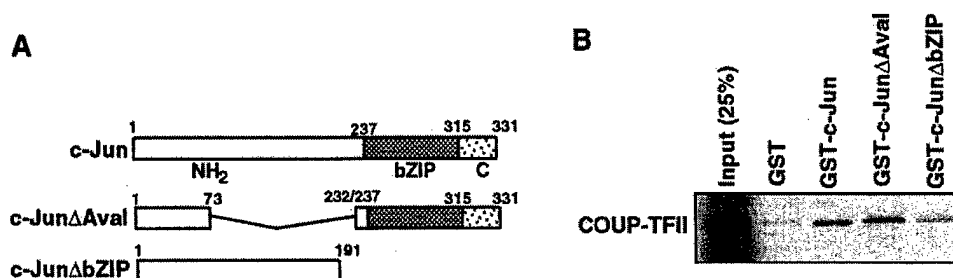


FIG. 5. The domain of c-Jun required for interaction with COUP-TF. A, schematic representation of c-Jun and its mutants. The amino terminus (NH₂), the basic region, the leucine zipper (bZIP), and the carboxyl terminus (C) are indicated. B, interaction between COUP-TF and c-Jun mutants. c-Jun and its mutant proteins were synthesized in bacteria using the pGEX-3X expression vector (Amersham Biosciences). GST-c-Jun and the mutant fusion proteins (GST-c-JunΔAval and GST-c-JunΔbZIP) were immobilized on the glutathione-Sepharose beads. As a control, the same amount of glutathione transferase was immobilized on the beads. *In vitro* translated ³⁵S-labeled COUP-TFII was then mixed with the beads. After extensive washing, bound proteins were analyzed by SDS-PAGE. Input proteins are shown for comparison.

GST pull-down assay (Fig. 4B). Cotransfection of the c-Jun expression vector alone induced reporter transcription (Fig. 4A), whereas cotransfection with one of the C-terminal deletion mutants COUP-TFIIΔ7, COUP-TFIIΔ30, or COUP-TFIIΔ108 strongly inhibited c-Jun-induced reporter activity, as was observed using the wild-type COUP-TFII. N-terminal domain deletion mutants, such as COUP-TFIIΔN25, COUP-TFIIΔN50, and COUP-TFIIΔN75, also retained the inhibitory effect on AP-1 activity by COUP-TFII. In contrast, partial (COUP-TFIIΔN125) or complete deletion of the DNA-binding domain (COUP-TFIIΔDBD) abrogated anti-c-Jun activity completely. In the GST pull-down assay, COUP-TFII mutants that effectively suppressed AP-1 activity, such as COUP-TFIIΔ7, COUP-TFIIΔ30, COUP-TFIIΔ108, COUP-TFIIΔN25, COUP-TFIIΔN50, and COUP-TFIIΔN75, were pulled down by GST-c-Jun protein, whereas COUP-TFIIΔDBD, which failed to suppress AP-1 activity, was not (Fig. 4B). Thus, inhibition of AP-1 transcriptional activity by the COUP-TFII mutants correlated with their ability to interact with c-Jun. These observations further suggest that a direct c-Jun/COUP-TF interaction accounts for the inhibition of AP-1 activity by COUP-TF and that the DBD of COUP-TF is essential.

We also identified the region of c-Jun required for COUP-TF interaction (Fig. 5). Deletion of amino acids 73–232 from c-Jun (c-JunΔAval) did not affect its ability to pull down COUP-TFII. In contrast, deletion of the C-terminal domain of c-Jun from amino acid 191 to 331 (c-JunΔbZIP), which encompasses the leucine-zipper region and basic region, completely abolished its ability to pull down COUP-TFII. Thus, the C-terminal domain, but not the N-terminal domain, of c-Jun is responsible for COUP-TF interaction.

Stable Expression of COUP-TF in COUP-TF-negative MDA-MB231 Cells Restores the Ability of *trans*-RA to Inhibit AP-1 Activity—To further examine the role of COUP-TF in *trans*-RA-induced inhibition of AP-1 activity, we analyzed the effect of *trans*-RA on TPA-induced c-Jun expression in MDA-MB231 breast cancer cells and MDA-MB231 cells stably transfected with COUP-TF (MB231/COUP#16) (35). Treatment of both cell lines with TPA strongly induced c-Jun expression as determined by Northern blotting (Fig. 6). Induction of c-Jun expression by TPA was probably due to an AP-1-binding site in the c-Jun promoter (7). Treatment with *trans*-RA did not affect basal c-Jun expression in both lines and did not inhibit TPA-induced c-Jun expression in MDA-MB231 cells. However, pretreatment of MB231/COUP#16 cells with *trans*-RA completely abolished TPA-induced c-Jun expression. These results demonstrate that COUP-TF expression is required for inhibition of AP-1 activity in MDA-MB231 cells by *trans*-RA.

Stable Expression of COUP-TF Does Not Lead to *trans*-RA-Dependent Inhibition of AP-1 Activity in HT-1376 Cells Lacking

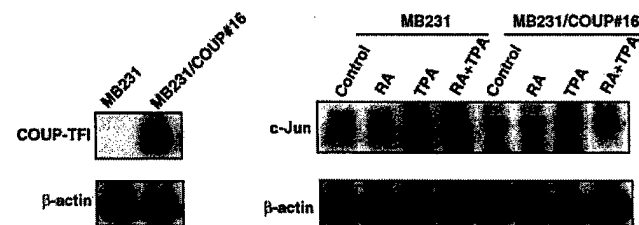


FIG. 6. Stable expression of COUP-TF in COUP-TF-negative, RAR-positive MDA-MB231 cells restores the ability of *trans*-RA to inhibit AP-1 activity. Total RNAs prepared from MDA-MB-231 or MDA-MB231 cells stably expressing COUP-TF (MB231/COUP#16) (35) treated with or without the indicated agents were analyzed for the expression of c-Jun. The expression of β-actin is shown to confirm the similar loading of RNA in each lane.

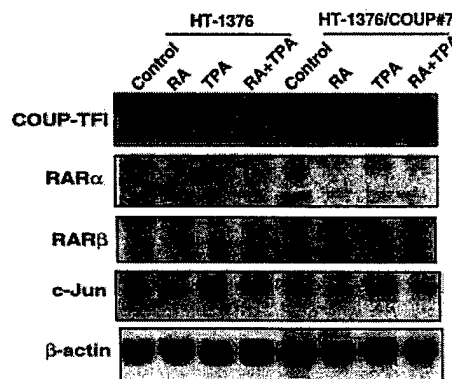


FIG. 7. Stable expression of COUP-TF in COUP-TF and RAR-negative HT-1376 cells does not lead to *trans*-RA-dependent inhibition of AP-1 activity. Total RNAs prepared from HT-1376 or HT-1376 cells stably expressing COUP-TF (HT-1376/COUP#7) treated with or without the indicated agents were analyzed for the expression of c-Jun, RARα, and RARβ. The expression of β-actin is shown to confirm similar loading of RNA in each lane.

RARα/β Expression—We also evaluated whether stable expression of COUP-TF affected TPA-induced c-Jun expression in HT-1376 bladder cancer cells (Fig. 7). The stable clone, HT-1376/COUP#7, which expresses high levels of COUP-TF, was analyzed for the effect of *trans*-RA on TPA activity (Fig. 7). About a 4-fold induction of c-Jun expression by TPA was observed in HT-1376 cells. However, the induction of c-Jun expression was largely reduced in HT-1376/COUP#7 cells, with only about a 2-fold induction. Interestingly, the basal level of c-Jun expression was also slightly reduced in HT-1376/COUP#7 cells. Despite the reduction of c-Jun expression in the absence or presence of TPA, *trans*-RA failed to inhibit basal and TPA-induced c-Jun expression in both wild-type and

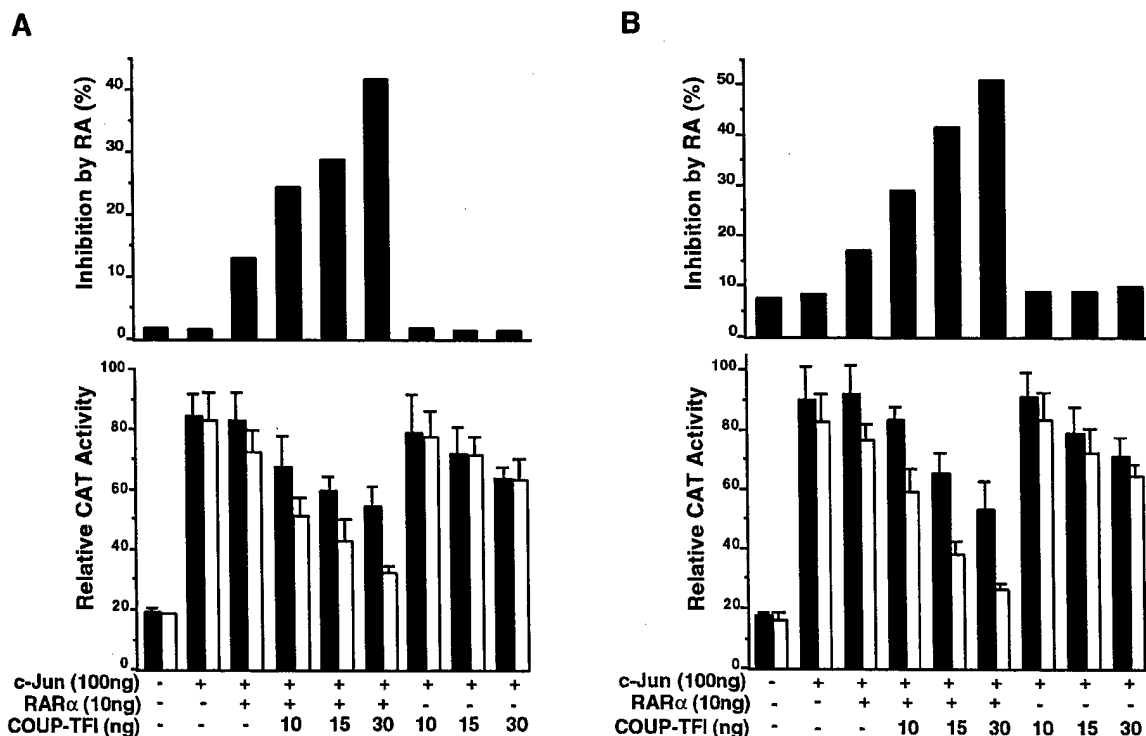


FIG. 8. Effect of COUP-TF and RAR α coexpression on antagonism of AP-1 activity by *trans*-RA. The -73Col-CAT reporter was transfected together with c-Jun alone or together with COUP-TF and/or RAR α expression vectors into HT-1376 (A) or HeLa (B) cells. After transfection, the cells were incubated in medium containing 0.5% FCS for 24 h and then treated with or without *trans*-RA. After 12 h, the cells were harvested, and CAT activity was determined. The top panels show the percentage of inhibition of c-Jun activity by *trans*-RA based on data shown in the bottom panels, in which filled bars represent control and open bars represent *trans*-RA treatment.

COUP-TF-expressing cells. Thus, COUP-TF expression is not sufficient to confer *trans*-RA-induced anti-AP-1 activity in HT-1376 cells.

RAR Expression Is Required for COUP-TF to Facilitate Antagonism of AP-1 Activity by *trans*-RA—Our observations that transient (Fig. 1B) or stable (Fig. 7) expression of COUP-TF in HT-1376 cells failed to modulate antagonism of AP-1 activity by *trans*-RA led us to investigate RAR expression. Unlike MDA-MB231 cells that express RAR α (48) and RAR β when COUP-TF is expressed (35), HT-1376 cells did not express detectable RAR α or RAR β (Fig. 7), although RAR γ was expressed (data not shown). This result suggested that modulation of *trans*-RA-induced anti-AP-1 activity by COUP-TF might require RAR α or RAR β . The effect of RAR α expression on the ability of COUP-TF to regulate *trans*-RA activity was then examined using transient transfection in HT-1376 cells. The -73Col-CAT reporter was transfected into HT-1376 cells with or without c-Jun and COUP-TF and/or RAR α vector. Transfected cells were treated with or without *trans*-RA. As shown in Fig. 8A, transfected COUP-TF repressed AP-1 activity in the absence of *trans*-RA. In contrast, transfected RAR α (10 ng) led to inhibition of AP-1 activity in a *trans*-RA dependent manner. The addition of COUP-TF significantly enhanced the *trans*-RA-induced inhibition of AP-1 activity by RAR α (Fig. 8A). Similar results were observed using HeLa cells (Fig. 8B). Thus, efficient inhibition of AP-1 activity by *trans*-RA activity by COUP-TF requires both COUP-TF and RAR α .

DISCUSSION

Retinoids are effective growth inhibitors of cancer cells. Inhibition of AP-1 activity has been proposed as one mechanism by which retinoids exert their anticancer effects (6). Despite extensive studies in the last few years, how retinoids specifically antagonize AP-1 activity remains largely unknown. Here, we provide evidence that COUP-TF is involved in regulating

the antagonism of AP-1 transactivational activity by *trans*-RA. COUP-TF, by physically interacting with c-Jun, inhibits AP-1 DNA binding and transactivation and is required for efficient inhibition of AP-1 activity by liganded RARs. Our results suggest that COUP-TF plays a role in the cross-talk between retinoid and AP-1 signaling pathways.

We recently reported that the expression of COUP-TF positively correlates with the inhibition of the growth of various cancer cell lines by *trans*-RA and that COUP-TF is underexpressed in many *trans*-RA-resistant cancer cell lines (35). Stable expression of COUP-TF in COUP-TF-negative cancer cells restores their sensitivity to *trans*-RA, demonstrating that COUP-TF can mediate anticancer effects of *trans*-RA (35). By studying anti-AP-1 activity of *trans*-RA in various cancer cell lines, we found a close correlation between COUP-TF expression and the anti-AP-1 activity of *trans*-RA (Fig. 1). In COUP-TF-positive ZR-75-1, T-47D, and Calu-6 cancer cell lines (35), *trans*-RA strongly inhibited the ability of TPA to activate transcription of the collagenase promoter, whereas in COUP-TF-negative MDA-MB231, H292, and HT-1376 cell lines, *trans*-RA failed to suppress TPA activity (Fig. 1). This finding is consistent with a previous study showing that *trans*-RA effectively inhibited AP-1 activity in ZR-75-1 and T-47D cells but not in MDA-MB231 cells (49). The requirement of COUP-TF in *trans*-RA-mediated AP-1 inhibition was further demonstrated by our findings that transient expression of COUP-TF in COUP-TF-negative cells (Fig. 1B) restored the ability of *trans*-RA to inhibit TPA-induced AP-1 transactivation and that stable expression of COUP-TF in MDA-MB231 cells enabled *trans*-RA to inhibit TPA-induced c-Jun expression (Fig. 6).

We have reported that COUP-TF expression contributes to growth inhibition by *trans*-RA in cancer cells because COUP-TF on binding to the RAR β promoter induces RAR β expression (35). RAR β is reported to be a potent AP-1 inhibitor

(24), suggesting that COUP-TF-induced RAR β probably contributes to the inhibition of AP-1 activity (Fig. 6). Our observations that COUP-TF can effectively interact with c-Jun *in vitro* (Figs. 3 and 4) and inhibit AP-1 transcriptional activity on transient transfection (Fig. 2) also suggest that COUP-TF is directly involved in antagonizing AP-1 activity.

Inhibition of AP-1 activity appears to be a common characteristic of nuclear receptors, as has been demonstrated for the progesterone, estrogen, androgen, thyroid hormone, glucocorticoid, and retinoid receptors, which can functionally interact with the AP-1 complex. This study reveals that the orphan receptor COUP-TF behaves similarly. The molecular basis of the interaction between nuclear receptors and AP-1 pathways remains largely unknown. Models proposed include direct protein-protein interaction (8, 9, 13–15), inhibition of Jun N-terminal kinase activity (31), and competition for CBP (32). Our results of GST pull-down assays (Figs. 4B and 5) and mutation analysis (Figs. 4 and 5) support the first model. The requirement for the COUP-TF DBD for c-Jun binding is similar to observations that the DBDs of glucocorticoid receptor (14, 15) and RAR (47) are essential for their interaction with AP-1. Interestingly, the *in vitro* DNA-binding study showed that COUP-TF is a more effective inhibitor of AP-1 binding than RAR α (Fig. 3) on the basis of the levels of each receptor to exert this effect.

In contrast to inhibition of AP-1 activity by other receptors that require their respective ligands, COUP-TF effectively inhibited AP-1 transactivation in the absence of any ligand (Fig. 2). Such a ligand-independent inhibition of AP-1 activity may restrict expression levels of AP-1-responsive genes in cancer cells (Figs. 6 and 7). Interestingly, we observed that COUP-TF expression potentiates antagonism of AP-1 activity by *trans*-RA. However, regulation of *trans*-RA activity by COUP-TF appears to require RAR α or RAR β , because transiently transfected COUP-TF was unable to confer anti-AP-1 activity to *trans*-RA in HT-1376 cells lacking RAR α or - β (Fig. 8A) or HeLa cells (Fig. 8B) unless RAR α was cotransfected. In addition, stable expression of COUP-TF restored *trans*-RA-induced anti-AP-1 activity in MDA-MB231 cells (Fig. 6) having RARs (48) but not in HT-1376 cells lacking RAR α and - β (Fig. 7). Thus, expression of both RAR α/β and COUP-TF is required for optimal inhibition of AP-1 activity by *trans*-RA.

Although we do not fully understand how COUP-TF and RAR α coexpression maximizes inhibition of AP-1 activity by *trans*-RA, our finding of physical interaction between COUP-TF and RAR α (35, 50) suggests its involvement in this process. Based on our observation that COUP-TF more effectively inhibits c-Jun binding to DNA than RAR α (Fig. 3), it is tempting to speculate that COUP-TF, with its ability to interact with both RAR (35, 50) and c-Jun (Fig. 3), may function as a bridging factor to mediate RAR/AP-1 interaction. In this model, the COUP-TF/RAR heterodimer would strongly interact with AP-1 in the presence of *trans*-RA to inhibit the DNA binding of AP-1 and its transactivation. Similarly, COUP-TF, by interacting with c-Jun, might also potentiate the inhibition of RAR α/β transactivation by AP-1. Unfortunately, perhaps due to rapid formation of the COUP-TF homodimer after COUP-TF expression *in vitro* (40, 51), we were unsuccessful in demonstrating the interaction between RAR/COUP-TF heterodimer and c-Jun in gel shift or GST pull-down assays (data not shown). Another possible mechanism for inhibiting AP-1 transactivation by COUP-TF may involve its recruiting of a transcriptional corepressor. COUP-TF has been widely considered as a potent negative transcriptional regulator (40–43) due to its effective interaction with corepressors, such as N-CoR

and SMRT (52). The interaction between COUP-TF and both RAR and AP-1 may recruit transcriptional corepressors to the complex, thereby enhancing the mutual repression of AP-1 and RAR transcriptional activity. Interestingly, COUP-TF interacts with a variety of nuclear receptors, including RXR (41), TR (40, 50), estrogen receptor (53, 54), and peroxisome proliferator-activated receptor (55). It remains to be determined whether and how COUP-TF is involved in modulating the anti-AP-1 activity by their ligands.

Acknowledgment—We thank Dr. Marcia I. Dawson for reviewing the manuscript.

REFERENCES

- Lotan, R. (1980) *Biochim. Biophys. Acta* **605**, 33–91
- Gudas, L. J., Sporn, M. B., and Roberts, A. B. (1994) in *The Retinoids*, 2nd Ed. (Sporn, M. B., Roberts, A. B., and Goodman, D. S., eds) pp. 443–520, Raven Press, New York
- Mangelsdorf, D. J., and Evans, R. M. (1995) *Cell* **83**, 841–850
- Kastner, P., Mark, M., and Chambon, P. (1995) *Cell* **83**, 859–869
- Zhang, X. K., and Pfahl, M. (1993) *Receptor* **3**, 183–191
- Pfahl, M. (1993) *Endocr. Rev.* **14**, 651–658
- Angel, P., and Karin, M. (1991) *Biochim. Biophys. Acta* **1072**, 129–157
- Yang-Yen, H. F., Zhang, X. K., Graupner, G., Tzukerman, M., Sakamoto, B., Karin, M., and Pfahl, M. (1991) *New Biol.* **3**, 1206–1219
- Schule, R., Rangarajan, P., Yang, N., Klier, S., Ransone, L. J., Bolado, J., Verma, I. M., and Evans, R. M. (1991) *Proc. Natl. Acad. Sci. U. S. A.* **88**, 6092–6096
- Nicholson, R. C., Mader, S., Nagpal, S., Leid, M., Rochette-Egly, C., and Chambon, P. (1990) *EMBO J.* **9**, 4443–4454
- Yang, L., Kim, H. T., Munoz-Medellin, D., Reddy, P., and Brown, P. H. (1997) *Cancer Res.* **57**, 4652–4661
- Soprano, D. R., Chen, L. X., Wu, S., Donigan, A. M., Borghaei, R. C., and Soprano, K. J. (1996) *Oncogene* **12**, 577–584
- Jonat, C., Rahmsdorf, H. J., Park, K. K., Cato, A. C., Gebel, S., Ponta, H., and Herrlich, P. (1990) *Cell* **62**, 1189–1204
- Yang-Yen, H. F., Chambard, J. C., Sun, Y. L., Smeal, T., Schmidt, T. J., Drouin, J., and Karin, M. (1990) *Cell* **62**, 1205–1215
- Schule, R., Rangarajan, P., Klier, S., Ransone, L. J., Bolado, J., Yang, N., Verma, I. M., and Evans, R. M. (1990) *Cell* **62**, 1217–1226
- Zhang, X. K., Dong, J. M., and Chiu, J. F. (1991) *J. Biol. Chem.* **266**, 8248–8254
- Zhang, X. K., Wills, K. N., Husmann, M., Hermann, T., and Pfahl, M. (1991) *Mol. Cell. Biol.* **11**, 6016–6025
- Shemshedini, L., Knauthe, R., Sassone-Corsi, P., Pornon, A., and Gronemeyer, H. (1991) *EMBO J.* **10**, 3839–3849
- Teyssier, C., Belguise, K., Galtier, F., and Chabos, D. (2001) *J. Biol. Chem.* **276**, 36361–36369
- Crawford, H. C., and Matrisian, L. M. (1996) *Enzyme Protein* **49**, 20–37
- Lee, H. Y., Dawson, M. I., Walsh, G. L., Nesbitt, J. C., Eckert, R. L., Fuchs, E., Hong, W. K., Lotan, R., and Kurie, J. M. (1996) *Cell Growth Differ.* **7**, 997–1004
- Agadir, A., Shealy, Y. F., Hill, D. L., and Zhang, X. (1997) *Cancer Res.* **57**, 3444–3450
- Zhang, X. K., Liu, Y., and Lee, M. O. (1996) *Mutat. Res.* **350**, 267–277
- Lin, F., Xiao, D., Kolluri, S. K., and Zhang, X. (2000) *Cancer Res.* **60**, 3271–3280
- Fanjul, A., Dawson, M. I., Hobbs, P. D., Jong, L., Cameron, J. F., Harlev, E., Graupner, G., Lu, X. P., and Pfahl, M. (1994) *Nature* **372**, 107–111
- Chen, J. Y., Penco, S., Ostrowski, J., Balaguer, P., Pons, M., Starrett, J. E., Reczek, P., Chambon, P., and Gronemeyer, H. (1995) *EMBO J.* **14**, 1187–1197
- Yang, L., Munoz-Medellin, D., Kim, H. T., Ostrowski, J., Reczek, P., and Brown, P. H. (1999) *Breast Cancer Res. Treat.* **56**, 277–291
- Li, J. J., Dong, Z., Dawson, M. I., and Colburn, N. H. (1996) *Cancer Res.* **56**, 483–489
- Huang, C., Ma, W. Y., Dawson, M. I., Rincon, M., Flavell, R. A., and Dong, Z. (1997) *Proc. Natl. Acad. Sci. U. S. A.* **94**, 5826–5830
- Konig, H., Ponta, H., Rahmsdorf, H. J., and Herrlich, P. (1992) *EMBO J.* **11**, 2241–2246
- Caelles, C., Gonzalez-Sancho, J. M., and Munoz, A. (1997) *Genes Dev.* **11**, 3351–3364
- Kamei, Y., Xu, L., Heinzel, T., Torchia, J., Kurokawa, R., Glass, B., Lin, S. C., Heyman, R. A., Rose, D. W., Glass, C. K., and Rosenfeld, M. G. (1996) *Cell* **85**, 403–414
- DiSepio, D., Sutter, M., Johnson, A. T., Chandraratna, R. A., and Nagpal, S. (1999) *Mol. Cell. Biol. Res. Commun.* **1**, 7–13
- Wu, Q., Li, Y., Liu, R., Agadir, A., Lee, M. O., Liu, Y., and Zhang, X. (1997) *EMBO J.* **16**, 1656–1669
- Lin, B., Chen, G. Q., Xiao, D., Kolluri, S. K., Cao, X., Su, H., and Zhang, X. K. (2000) *Mol. Cell. Biol.* **20**, 957–970
- Miyajima, N., Kadowaki, Y., Fukushima, S., Shimizu, S., Semba, K., Yamanashi, Y., Matsubara, K., Toyoshima, K., and Yamamoto, T. (1988) *Nucleic Acids Res.* **16**, 11057–11074
- Wang, L. H., Tsai, S. Y., Cook, R. G., Beattie, W. G., Tsai, M. J., and O'Malley, B. W. (1989) *Nature* **340**, 163–166
- Ladiaz, J. A., and Karathanasis, S. K. (1991) *Science* **251**, 561–565
- Qiu, Y., Krishnan, V., Pereira, F. A., Tsai, S. Y., and Tsai, M. J. (1996) *J.*

- Steroid Biochem. Mol. Biol.* **56**, 81-85
40. Tran, P., Zhang, X. K., Salbert, G., Hermann, T., Lehmann, J. M., and Pfahl, M. (1992) *Mol. Cell. Biol.* **12**, 4666-4676
41. Kliewer, S. A., Umesono, K., Heyman, R. A., Mangelsdorf, D. J., Dyck, J. A., and Evans, R. M. (1992) *Proc. Natl. Acad. Sci. U. S. A.* **89**, 1448-1452
42. Cooney, A. J., Tsai, S. Y., O'Malley, B. W., and Tsai, M. J. (1992) *Mol. Cell. Biol.* **12**, 4153-4163
43. Widom, R. L., Rhee, M., and Karathanasis, S. K. (1992) *Mol. Cell. Biol.* **12**, 3380-3389
44. Zhang, X. K., Hoffmann, B., Tran, P. B., Graupner, G., and Pfahl, M. (1992) *Nature* **355**, 441-446
45. Wu, Q., Dawson, M. I., Zheng, Y., Hobbs, P. D., Agadir, A., Jong, L., Li, Y., Liu, R., Lin, B., and Zhang, X. K. (1997) *Mol. Cell. Biol.* **17**, 6598-6608
46. Li, Y., Lin, B., Agadir, A., Liu, R., Dawson, M. I., Reed, J. C., Fontana, J. A., Bost, F., Hobbs, P. D., Zheng, Y., Chen, G. Q., Shroot, B., Mercola, D., and Zhang, X. K. (1998) *Mol. Cell. Biol.* **18**, 4719-4731
47. Zhou, X. F., Shen, X. Q., and Shemshedini, L. (1999) *Mol. Endocrinol.* **13**, 276-285
48. Liu, Y., Lee, M. O., Wang, H. G., Li, Y., Hashimoto, Y., Klaus, M., Reed, J. C., and Zhang, X. (1996) *Mol. Cell. Biol.* **16**, 1138-1149
49. van der Burg, B., Slager-Davidov, R., van der Leede, B. M., de Laat, S. W., and van der Saag, P. T. (1995) *Mol. Cell. Endocrinol.* **112**, 143-152
50. Berrodrin, T. J., Marks, M. S., Ozato, K., Linney, E., and Lazar, M. A. (1992) *Mol. Endocrinol.* **6**, 1468-1478
51. Butler, A. J., and Parker, M. G. (1995) *Nucleic Acids Res.* **23**, 4143-4150
52. Shibata, H., Nawaz, Z., Tsai, S. Y., O'Malley, B. W., and Tsai, M. J. (1997) *Mol. Endocrinol.* **11**, 714-724
53. Klinge, C. M., Silver, B. F., Driscoll, M. D., Sathya, G., Bambara, R. A., and Hilf, R. (1997) *J. Biol. Chem.* **272**, 31465-31474
54. Klinge, C. M. (1999) *J. Steroid Biochem. Mol. Biol.* **71**, 1-19
55. Robinson, C. E., Wu, X., Nawaz, Z., Onate, S. A., and Gimble, J. M. (1999) *Endocrinology* **140**, 1586-1593

Regulation of Retinoic Acid Receptor β Expression by Peroxisome Proliferator-activated Receptor γ Ligands in Cancer Cells¹

Sharon Y. James, Feng Lin, Siva Kumar Kolluri, Marcia I. Dawson, and Xiao-kun Zhang²

Cancer Center, The Burnham Institute, La Jolla, California 92037

ABSTRACT

The peroxisome proliferator-activated receptor γ (PPAR γ) is a nuclear receptor family member that can form a heterodimeric complex with retinoid X receptor (RXR) and initiate transcription of target genes. In this study, we have examined the effects of the PPAR γ ligand ciglitazone and the RXR ligand SR11237 on growth and induction of retinoic acid receptor (RAR) β expression in breast and lung cancer cells. Our results demonstrated that ciglitazone and SR11237 cooperatively inhibited the growth of ZR-75-1 and T-47D breast cancer and Calu-6 lung cancer cells. Gel shift analysis indicated that PPAR γ , in the presence of RXR, formed a strong complex with a retinoic acid response element (β retinoic acid response element) in the RAR β promoter. In reporter gene assays, RXR ligands and ciglitazone, but not the PPAR γ ligand 15d-PGJ₂, cooperatively promoted the transcriptional activity of the β retinoic acid response element. Ciglitazone, but not 15d-PGJ₂, strongly induced RAR β expression in human breast and lung cancer cell lines when used together with SR11237. The induction of RAR β expression by the ciglitazone and SR11237 combination was diminished by a PPAR γ -selective antagonist, bisphenol A diglycidyl ether. All-*trans*-retinoic acid or the combination of ciglitazone and SR11237 was able to induce RAR β in all-*trans*-retinoic acid-resistant MDA-MB-231 breast cancer cells only when the orphan receptor chick ovalbumin upstream promoter transcription factor was expressed, or in the presence of the histone deacetylase inhibitor trichostatin A. These studies indicate the existence of a novel RAR β -mediated signaling pathway of PPAR γ action, which may provide a molecular basis for developing novel therapies involving RXR and PPAR γ ligands in potentiating antitumor responses.

INTRODUCTION

PPAR γ ³ is a ligand-activated transcription factor belonging to the steroid/thyroid receptor superfamily, which plays a critical role in the control of adipogenesis (1–4). Specific ligands of PPAR γ , including the thiazolidinedione class of antidiabetic agents, the prostanoid 15d-PGJ₂, and certain polyunsaturated fatty acids, have been identified (5, 6). PPAR γ expression is not limited to adipocytes because activation of PPAR γ by its ligands has been shown to promote growth inhibition, differentiation, and/or apoptosis of various cancer cells (5, 7–9), including breast (10–18) and non-small cell lung carcinoma tissues (19–22).

Received 10/28/02; accepted 4/24/03.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

¹ S. J. and S. K. K. are supported by Postdoctoral Research Fellowships from the California Breast Cancer Research Program (BCRP 7FB-0062) and the United States Army Medical Research Program (DAMD17-00-1-0173), respectively. M. I. D. and X.-k. Z. are supported by National Cancer Institute Grant PO1 CA51993. X.-k. Z. is also supported by grants from the NIH (5R01 CA 60988-09 and 7P01 CA 87000-01A1) and the United States Army Medical Research Program (DAMD17-00-1-0172).

² To whom requests for reprints should be addressed, at Cancer Center, The Burnham Institute, 10901 North Torrey Pines Road, La Jolla, CA 92037. Phone: (858) 646-3141; Fax: (858) 646-3195; E-mail: xzhang@burnham-inst.org.

³ The abbreviations used are: PPAR γ , peroxisome proliferator-activated receptor γ ; 15d-PGJ₂, 15-deoxy- $\Delta^{12,14}$ prostaglandin J₂; ATRA, all-*trans*-retinoic acid; β RARE, β retinoic acid response element; BADGE, bisphenol A diglycidyl ether; COUP-TF, chick ovalbumin upstream promoter transcription factor; DR, direct repeat; HDAC, histone deacetylase; PPRE, peroxisome proliferator response element; RAR, retinoic acid receptor; RXR, retinoid X receptor; TSA, trichostatin A; CAT, chloramphenicol transferase; β -gal, β -galactosidase; MTS, 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium; BrdUrd, 5-bromo-2'-deoxyuridine; tk, thymidine kinase.

Retinoids, comprising the native and synthetic derivatives of vitamin A, are promising agents for the prevention and treatment of human cancers, including those of breast and lung (23, 24). The biological effects of retinoids are mainly mediated by their nuclear receptors, RAR and RXR, which each exist as α , β , and γ isoforms (1, 25, 26). PPAR γ heterodimerizes with RXR, and the resulting heterodimer binds strongly to its DNA-specific sequence, the PPRE (3, 4). Recent studies have shown that PPAR γ /RXR also binds to the estrogen response element (27). PPAR γ and RXR ligands have been found to cooperatively induce differentiation and apoptosis of breast and colon cancer cells through interaction with PPAR γ /RXR (17, 18, 23, 28).

RAR β plays a critical role in mediating the growth-inhibitory effects of retinoids in various cancer cells (29, 30). The aberrant expression or loss of RAR β in a variety of cancer cell lines suggests that decreased RAR β expression may contribute to retinoid resistance (31–38), implying that RAR β may act as a tumor suppressor. Regulation of RAR β gene expression is principally mediated by the retinoic acid response element (β RARE) in its promoter, to which RAR/RXR heterodimers bind strongly. Activation of RAR/RXR heterodimers is principally mediated via RAR, whereas RXR serves as a silent partner (25). The TR3/RXR heterodimer also binds to the β RARE, which is activated by RXR ligands (39). Thus, the β RARE can be transcriptionally controlled by various heterodimers and their ligands. Other factors also influence RAR β gene expression. The COUP-TF, which is not expressed in many cancer cells, is required for induction of RAR β by ATRA (40). Lack of RAR β expression in cancer cells has also been attributed to abnormal regulation of histone acetylation/deacetylation, which modulates chromatin structure and gene transcription, or hypermethylation of the RAR β promoter (41–45).

In the present study, we evaluated the growth-inhibitory effect of the PPAR γ ligand ciglitazone alone and in combination with the RXR ligand (retinoid) SR11237. Our results showed that ciglitazone and SR11237 cooperatively inhibited the growth and induced apoptosis of breast and lung cancer cell lines. In studying the possible underlying molecular mechanisms, we observed that PPAR γ could bind strongly to the β RARE as a PPAR γ /RXR heterodimer. The combination of RXR ligands with ciglitazone, but not 15d-PGJ₂, strongly activated the β RARE and induced RAR β expression in breast and lung cancer cells. The induction of RAR β expression by retinoids and ciglitazone was reduced by a PPAR γ antagonist, BADGE (46), indicating the involvement of PPAR γ /RXR heterodimers. Together, our results demonstrate that PPAR γ can bind to the β RARE as a PPAR γ /RXR heterodimer and that induction of RAR β may contribute to the anticancer effect of certain PPAR γ ligands.

MATERIALS AND METHODS

Reagents and Cell Lines. The RXR-selective retinoids SR11237, SR11246, and SR11345 were prepared as reported previously (47–49). ATRA and TSA were obtained from Sigma Chemicals (St. Louis, MO). The PPAR γ ligands 15d-PGJ₂, ciglitazone, and BADGE were obtained from Cayman Chemicals (Ann Arbor, MI). All reagents were dissolved in a 1:1 ratio of ethanol and DMSO and stored in amber containers at -20°C . Other analytical-grade reagents were obtained from Sigma Chemicals unless otherwise stated.

MDA-MB-231, T-47D, and ZR-75-1 cell lines were routinely maintained in DMEM (MDA-MB-231) or RPMI 1640 (T-47D and ZR-75-1), supplemented with 10% FCS, 100 units/ml penicillin, and 100 μ g/ml streptomycin (Irvine Scientific, Santa Ana, CA). Calu-6 were maintained in MEM Earle's Salt Medium (Irvine Scientific) supplemented with 10% FCS and antibiotics. For Western analysis studies, cells were cultured in their respective medium supplemented with 5% charcoal-treated FCS and antibiotics.

Plasmids. The PPAR γ expression vector was kindly provided by Dr. Mark Leid (Oregon State University, Corvallis, OR). Expression vectors for RXR α and reporter gene β RARE-tk-CAT have been described previously (26, 50).

Transfection Assays. MDA-MB-231 and ZR-75-1 cells were seeded at 2×10^5 cells/ml in 6-well plates for 16–24 h before transfection. Cells were transfected with β RARE-tk-CAT plasmid (200 ng), β -gal (200 ng) expression vector (pCH 110; Amersham Biosciences), and carrier DNA (pBluescript; Stratagene, La Jolla, CA) to a final concentration of 2000 ng total DNA/well using LipofectAMINE Plus reagent (Invitrogen, Carlsbad, CA). Cells were treated for 20 h with RXR and PPAR γ ligands. CAT activity was normalized with β -gal activity for transfection efficiency.

Western Blot Analysis. Treated cell cultures were subjected to Western blot analysis as described previously (51). Preblocked nitrocellulose membranes were incubated with 1 μ g/ml equivalent of anti-RAR β rabbit polyclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA). RAR β protein was detected by horseradish peroxidase-conjugated antirabbit secondary antibody (Amersham Biosciences), and specific bands were visualized by enhanced chemiluminescence (ECL; Amersham Biosciences). Equivalent loading of samples was determined by reprobing the nitrocellulose membrane with a mouse monoclonal antibody recognizing β -actin (Sigma Chemicals).

Gel Shift Analysis. *In vitro*-synthesized RXR and PPAR γ receptor proteins (Promega, Madison, WI) were incubated with RXR and PPAR γ ligands alone or in combination and treated with a rabbit polyclonal anti-PPAR γ antibody (Santa Cruz Biotechnology) for 20 min before the addition of 32 P-labeled β RARE oligonucleotide. Gel shift analysis was performed as described previously (35).

Cell Proliferation and Apoptosis Studies. Treated cells were trypsinized, pelleted by centrifugation at 2000 rpm for 5 min, resuspended in 1 ml of PBS, and fixed in 70% ice-cold ethanol. After two additional PBS washes, the cells were resuspended in PBS containing 50 μ g/ml propidium iodide (Sigma Chemicals) and 100 μ g/ml DNase-free RNase A (Roche Diagnostics, Indianapolis, IN). Cell suspensions were incubated for 30 min at 37°C with protection from light and analyzed using a FACScatter-Plus flow cytometer (Becton Dickinson, San Jose, CA).

To assess cell viability, cells were seeded at 1×10^3 cells/well in 96-well microtiter plates and treated with varying concentrations of SR11237 and ciglitazone, with medium and ligands replaced every 48 h. After treatment, 20 μ l of MTS/phenazine methosulfate solution (Promega) were added to each well, and incubation was continued for 2–4 h at 37°C in the dark. Absorbance (490 nm) was measured on a Bio-Rad 550 microplate reader.

BrdUrd Analysis. Treated cells were incubated with BrdUrd (20 μ M; Sigma Chemicals) for 2 h before harvesting of cells. After trypsinization and two PBS washes, cells were pelleted by centrifugation and permeabilized with 4% paraformaldehyde. After a 20-min incubation at room temperature, 0.1% saponin (Sigma Chemicals) was added to the cell suspension, and the incubation was continued for 10 min. The cells were then centrifuged, washed twice with PBS containing 0.1% saponin, and resuspended in PBS containing 30 μ g of DNase I (Roche Diagnostics). After a 1-h incubation with either an anti-BrdUrd fluorescent antibody or an isotope control antibody (Becton Dickinson), cells were given a final PBS wash before being analyzed using the FACScatter-Plus flow cytometer (Becton Dickinson).

Statistical Analysis. One-way ANOVA with the Dunnett's post test (GraphPad Prism software) was used to assess significance of treatments at the 5% level for growth inhibition studies.

RESULTS

Ciglitazone and SR11237 Cooperatively Inhibit the Growth of Cancer Cell Lines. Both retinoids and PPAR γ ligands are potent regulators of cancer cell growth. Because PPAR γ heterodimerizes with RXR (5, 52), we investigated the inhibitory effect of their

ligands, alone or in combination, on the growth of breast and lung cancer cells. Fig. 1A illustrates the effects of the retinoid SR11237 and the PPAR γ ligand ciglitazone on the growth of the hormone-dependent breast cancer cell lines ZR-75-1 and T-47D and the lung cancer cell line Calu-6. Cell proliferation was assessed by MTS assay after 8 (ZR-75-1 and Calu-6) or 10 (T-47D) days of treatment. All three cell lines have been reported to be sensitive to growth inhibition by ATRA (35, 53). Treatment of these cell lines with ATRA strongly inhibited their growth in a dose-dependent manner (Fig. 1A). SR11237 did not exhibit a significant effect on T-47D cell growth, whereas in ZR-75-1 and Calu-6 cells, the retinoid modestly inhibited cell growth when used at 1 μ M (10% and 22% for ZR-75-1 and Calu-6 cells, respectively; Fig. 1A). Ciglitazone (1 μ M) did not have any marked antiproliferative effects in these cancer cell lines. However, when SR11237 and ciglitazone were used in combination, cooperative effects on cell growth were observed in the three cell lines. In ZR-75-1 cells, a 2-fold growth-inhibitory effect was observed when using 1 μ M SR11237 and 1 μ M ciglitazone (10% with SR11237 versus 23% with the combination). In addition, cotreatment of ZR-75-1 cells with 1 μ M SR11237 and 10 μ M ciglitazone further enhanced growth inhibition by 47% (Fig. 1A, middle graph). Cotreatment of T-47D cells with SR11237 and ciglitazone resulted in 51% growth inhibition, compared with the antiproliferative effects of each ligand used alone, whereas an additive effect was observed in Calu-6 cells.

We also evaluated the antiproliferative effects of SR11237 and ciglitazone by BrdUrd incorporation. ZR-75-1 cells were treated for 72 h with SR11237 alone or in combination with ciglitazone. As shown in Fig. 1B, SR11237 or ciglitazone alone did not inhibit BrdUrd incorporation relative to control. In contrast, the combination decreased BrdUrd incorporation by 46%. Additional studies were performed to determine whether the combination induced apoptosis (Fig. 1C). ZR-75-1 cells were treated for 5 days with SR11237 alone or in the presence of ciglitazone. After treatment, cells were harvested and stained with propidium iodide for flow cytometry. Apoptotic cells were determined by measuring sub-G₁-phase cells. When the cells were treated with the combination of SR11237 and ciglitazone, 7.6% of the cells were apoptotic compared with control (Fig. 1C). Taken together, these results demonstrate that ciglitazone and a RXR ligand can cooperatively inhibit growth and induce apoptosis in cancer cells.

Induction of RAR β by a PPAR γ Ligand in Cancer Cell Lines. Induction of RAR β has been correlated with the growth-inhibitory and apoptosis-inducing effects of retinoids in breast and lung cancer cells (30, 35, 36). We then determined whether the antiproliferative effects observed using retinoids and PPAR γ ligands could be attributed to their ability to induce RAR β . Western analysis was used to investigate whether RXR and PPAR γ ligands were capable of regulating RAR β protein expression. ZR-75-1, T-47D, and Calu-6 cells were treated for 24 h with SR11237, in the absence or presence of ciglitazone. In these cell lines, ATRA readily induced RAR β , as reported previously (35), whereas ciglitazone did not show any effect (Fig. 2). SR11237 only weakly induced RAR β in all of the cell lines. However, cotreatment with SR11237 and ciglitazone resulted in a marked expression of RAR β protein. Together, these data demonstrate that ciglitazone strongly induces RAR β expression in combination with a RXR ligand.

Differential Effects of PPAR γ Ligands on RAR β Expression. Next, we examined whether other known PPAR γ ligands were able to induce RAR β . We evaluated the prostanoid 15d-PGJ₂ and rosiglitazone, two well-characterized PPAR γ ligands (5, 6). T-47D, ZR-75-1, and Calu-6 cells were first treated with 15d-PGJ₂ alone or with SR11237. Unlike SR11237 and ciglitazone cotreatment (Fig. 2), SR11237 and 15d-PGJ₂ did not induce RAR β (Fig. 3), thus highlight-

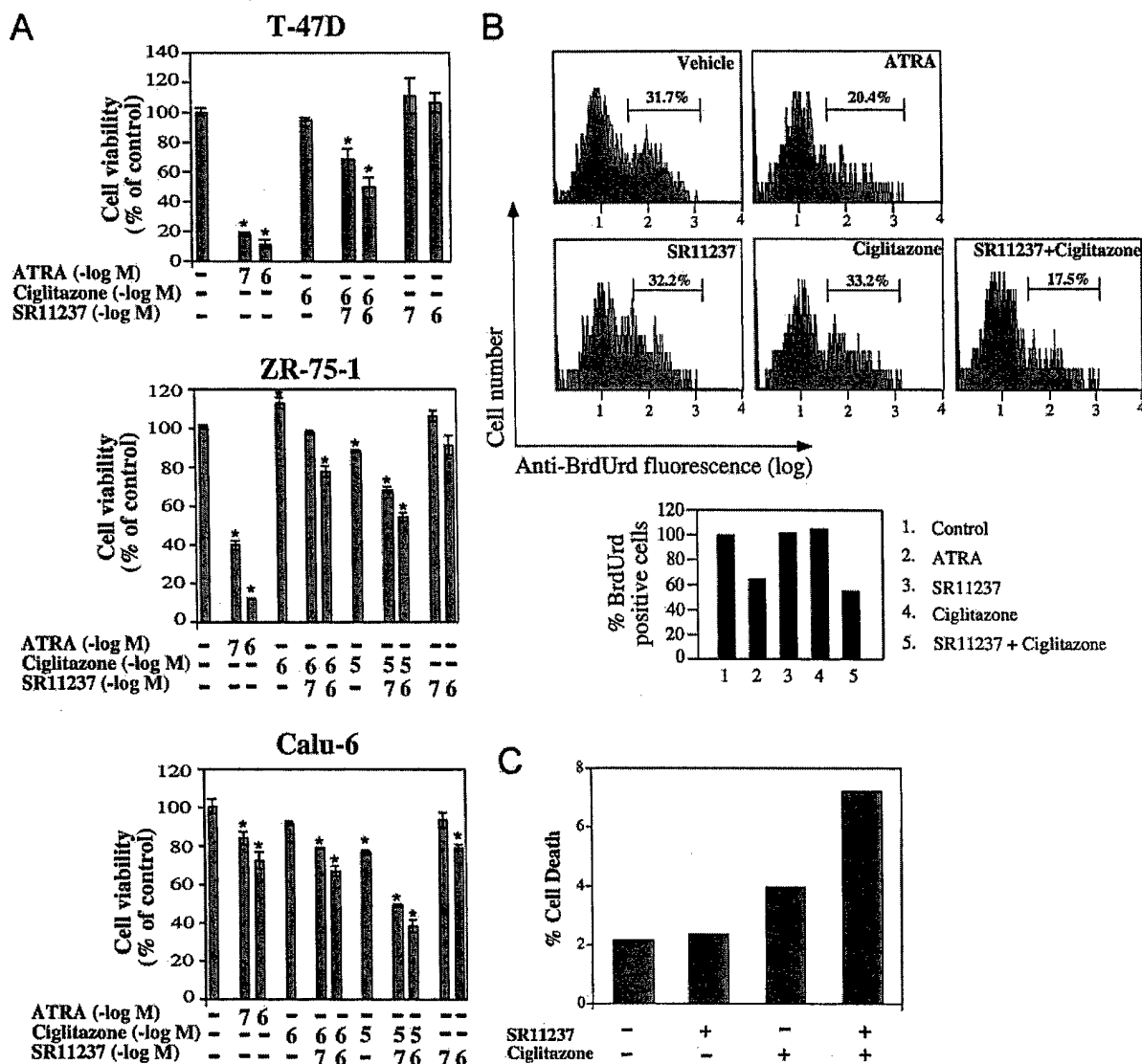


Fig. 1. PPAR γ ligand ciglitazone and RXR ligand SR11237 cooperatively inhibit growth and induce apoptosis of cancer cells. A, inhibition of cell proliferation by RXR and PPAR γ ligands. T-47D, ZR-75-1, and Calu-6 cells were treated for 10 (T-47D) or 8 (ZR-75-1 and Calu-6) days with the indicated concentrations of ATRA, SR11237, or ciglitazone. After treatment, inhibition of cell proliferation was assessed by MTS assay. Cell viability is expressed as a percentage of the control \pm SE ($n = 4$). *, $P < 0.05$. B, retinoid and ciglitazone cooperatively inhibit breast cancer cell proliferation. ZR-75-1 cells were treated for 72 h with ATRA alone ($1 \mu\text{M}$, as positive control) or with SR11237 ($1 \mu\text{M}$) or ciglitazone ($10 \mu\text{M}$), either alone or in combination. Cell proliferation was assessed by BrdUrd incorporation (top graphs). The data are presented as a histogram (bottom graph). C, induction of apoptosis by RXR and PPAR γ ligands. ZR-75-1 cells were treated for 5 days with SR11237 ($1 \mu\text{M}$) or ciglitazone ($10 \mu\text{M}$), either alone or in combination. Apoptotic cells were determined by measuring the sub-G $_1$ peak of cells using propidium iodide staining and flow cytometry. Results are representative of three separate experiments.

ing the differential effects of the PPAR γ ligands on modulating RAR β expression in combination with a RXR-selective retinoid. We also found that rosiglitazone alone or with SR11237 did not induce RAR β (Fig. 3). Thus, not all PPAR γ ligands are capable of inducing RAR β expression in cancer cells.

PPAR γ Mediates the Effects of Ciglitazone in Inducing RAR β .

To determine whether the induction of RAR β by ciglitazone (in combination with SR11237) was mediated by PPAR γ , we examined the effect of BADGE a PPAR γ antagonist. BADGE was shown to block the ability of PPAR γ ligands to activate the transcriptional and adipogenic function of PPAR γ (46). Fig. 4 illustrates that BADGE alone did not affect basal RAR β expression in T-47D cells. However, BADGE significantly attenuated ciglitazone/SR11237-induced RAR β . The effect of BADGE was specific because it did not inhibit ATRA-induced RAR β expression. Similar attenuation of ciglitazone/SR11237-induced RAR β by BADGE was observed in ZR-75-1 cells (data not shown). These data strongly suggest that induction of RAR β

expression by ciglitazone (in combination with SR11237) is mediated via activation of PPAR γ .

PPAR γ Binds to the β RARE as a PPAR γ /RXR Heterodimer.

The β RARE present in the RAR β promoter mediates transcriptional regulation of RAR β expression by RAR and RXR heterodimers (50, 54). To determine whether induction of RAR β by ciglitazone and SR11237 was also mediated via β RARE, we studied the possibility that PPAR γ was capable of interacting with the β RARE. *In vitro*-translated PPAR γ protein was analyzed for binding to the β RARE by the gel shift assay (Fig. 5A). PPAR γ alone did not exhibit clear binding to the β RARE. However, PPAR γ in combination with *in vitro*-synthesized RXR α protein produced a prominent complex. Binding was not affected when PPAR γ /RXR was preincubated with SR11237 or ciglitazone. However, when *in vitro*-translated RXR and PPAR γ were incubated with an anti-PPAR γ antibody, binding of the RXR/PPAR γ heterodimer to β RARE was attenuated, indicating that the PPAR γ antibody blocked the ability of PPAR γ to bind to β RARE

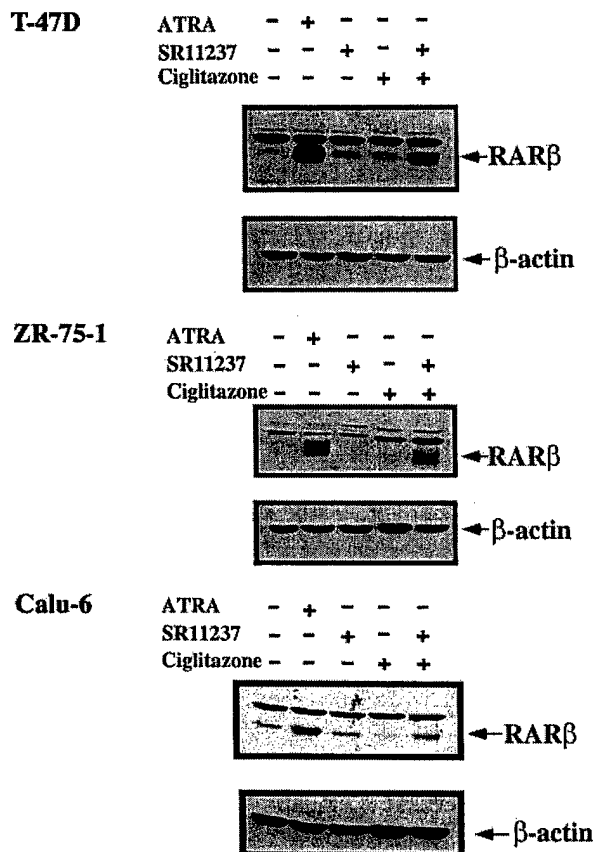


Fig. 2. Regulation of RAR β protein expression by RXR and PPAR γ ligands. T-47D, ZR-75-1, and Calu-6 cells were treated for 24 h with SR11237 (1 μ M) or ciglitazone (10 μ M), either alone or in combination. Cells were also treated with ATRA (1 μ M) as a positive control. Cell lysates were prepared, and RAR β protein expression was assessed by Western analysis (51).

(Fig. 5B). These data demonstrate that PPAR γ can bind to the β RARE as a PPAR γ /RXR heterodimer.

Differential Regulation of β RARE Transcriptional Activity by PPAR γ Ligands. The observation that PPAR γ /RXR binds to the β RARE suggested that it represented another RXR-containing heterodimeric complex that activates the β RARE. Therefore, we studied whether RXR and PPAR γ ligands could activate the β RARE. A reporter containing the CAT gene fused to the β RARE (50) was transiently transfected into MDA-MB-231 cells, which were treated with SR11237 alone or in combination with ciglitazone. As revealed in Fig. 6A, SR11237, but not ciglitazone, slightly induced β RARE transcriptional activity. Moreover, induction of β RARE was synergized by cotreatment with ciglitazone. We also examined whether the synergistic effect on activating β RARE could be extended to other PPAR γ and RXR ligands. Ciglitazone, but not 15d-PGJ₂ or rosiglitazone, strongly induced β RARE activity together with SR11237 (Fig. 6A). This correlated with the effects of these PPAR γ ligands on induction of RAR β protein expression (Figs. 2 and 3). Similar results were obtained using ZR-75-1 breast cancer cells (Fig. 6B, top graph). Thus, not all PPAR γ ligands that activate the PPARE similarly activate β RARE together with SR11237. We also examined the effect of the RXR ligands SR11246 and SR11345 on β RARE activity in ZR-75-1 cells (48, 49). Again, ciglitazone strikingly enhanced transcriptional activity of the β RARE induced by SR11246 and SR11345 (Fig. 6B).

Antagonistic Effects of Other PPAR γ Ligands on RAR β Expression. The differential effect of PPAR γ ligands on SR11237-induced RAR β expression (Figs. 2 and 3) prompted us to investigate whether PPAR γ ligands antagonized one another's activity. The ob-

servation that ciglitazone synergized with SR11237 in inducing RAR β expression suggested that binding of ciglitazone to PPAR γ and binding of SR11237 to RXR resulted in transactivation of the β RARE by PPAR γ /RXR heterodimers. In contrast, binding of PPAR γ with 15d-PGJ₂ may either not activate or suppress the transcriptional activity of PPAR γ /RXR on the β RARE. Therefore, we determined whether 15d-PGJ₂ interfered with PPAR γ activation by ciglitazone. Fig. 7 shows that in ZR-75-1 cells, 15d-PGJ₂ inhibited RAR β expression induced by the combination of SR11237 and ciglitazone. Thus, 15d-PGJ₂ acts as an antagonist of PPAR γ with respect to the induction of RAR β by ciglitazone.

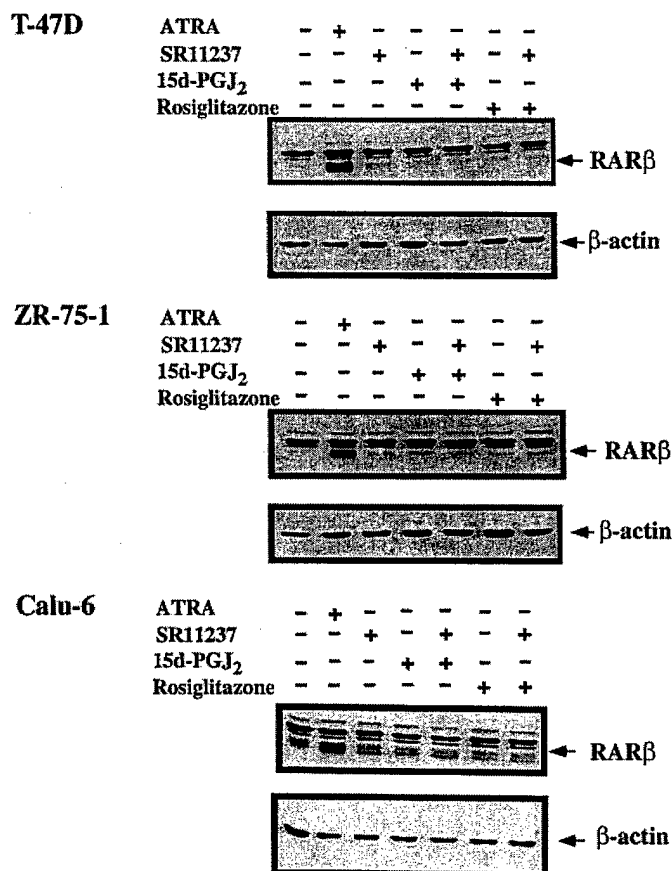


Fig. 3. Differential effects of PPAR γ ligands in inducing RAR β expression. T47D, ZR-75-1 and Calu-6 cells were treated for 24 h with SR11237 (1 μ M), 15d-PGJ₂ (5 μ M), or rosiglitazone (10 μ M), alone or with SR11237 + 15d-PGJ₂ or SR11237 + rosiglitazone in combination. Cells were also treated with ATRA (1 μ M), as a positive control. RAR β protein expression was assessed by Western analysis.

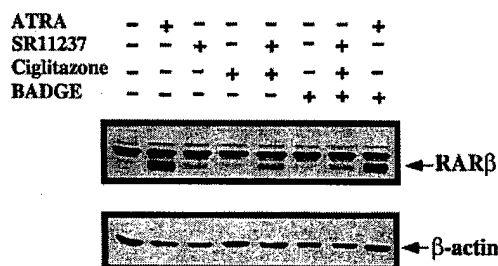


Fig. 4. A PPAR γ antagonist blocks induction of RAR β protein expression. T-47D cells were pretreated for 3 h with the PPAR γ antagonist BADGE (20 μ M), after which the cells were treated with SR11237 (1 μ M) or ciglitazone (10 μ M), alone or in combination. RAR β protein expression was determined by Western analysis.

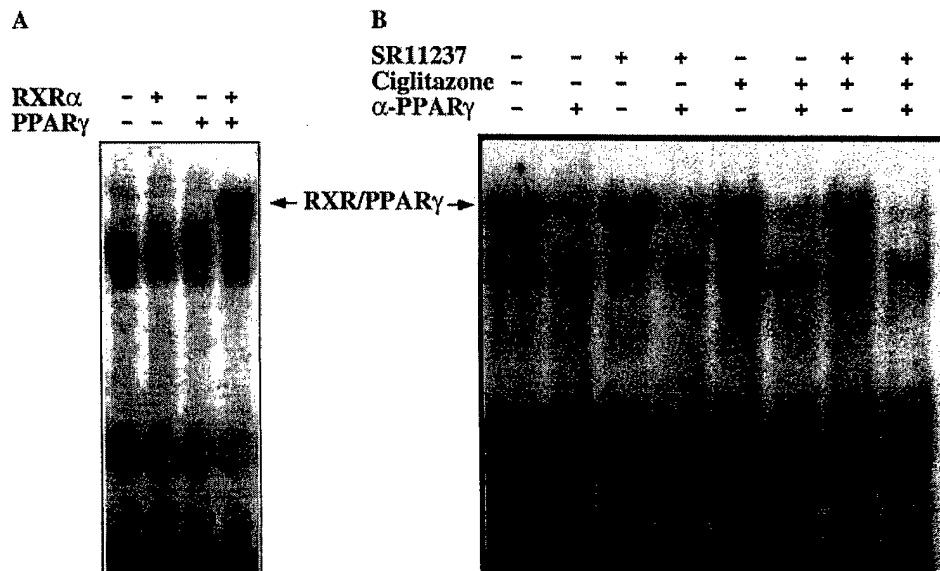


Fig. 5. RXR and PPAR γ bind to β RARE as a heterodimeric complex. **A**, *in vitro*-translated RXR and PPAR γ were incubated with 32 P-radiolabeled β RARE, either alone or in combination. The resulting reactions were then analyzed by gel shift analysis. **B**, *in vitro*-translated RXR α and PPAR γ were incubated with SR11237 (1 μ M) and/or ciglitazone (10 μ M), in the absence or presence of anti-PPAR γ rabbit polyclonal antibody (α -PPAR γ). After a further incubation with 32 P-radiolabeled β RARE, the reactions were analyzed by gel shift analysis.

Mechanisms of RAR β Induction in MDA-MB-231 Cells: Role of HDACs and the Orphan Receptor COUP-TF. ATRA does not induce RAR β expression in the estrogen-independent breast cancer cell line MDA-MB-231 (35, 41–43, 55, 56). Therefore, we examined whether SR11237 and ciglitazone could induce RAR β in these cells. In agreement with previous observations (35), treatment of MDA-MB-231 cells with ATRA did not induce RAR β expression (Fig. 8A). SR11237 and ciglitazone, either alone or in combination, did not induce RAR β expression, despite the expression of RXR α and PPAR γ (data not shown). Histone acetylation and DNA methylation have been reported to contribute to silencing RAR β gene expression in the MDA-MB-231 cell line (41, 44). HDAC inhibitors, such as TSA, have been shown to suppress the effects of corepressors (41). Therefore, to determine whether inhibitors of histone deacetylation played a role in augmenting induction of RAR β by RXR and PPAR γ ligands, MDA-MB-231 cells were treated with SR11237, alone or in combination with ciglitazone and TSA. Cotreatment of cells with ATRA and TSA readily induced RAR β protein expression (Fig. 8A). Cells cotreated with SR11237, ciglitazone, and TSA also induced RAR β . These results suggest that histone deacetylation may represent a mechanism through which RAR β expression is suppressed in MDA-MB-231 cancer cells.

We reported previously that the orphan receptor COUP-TF is required for RAR β expression by ATRA and is not expressed in MDA-MB-231 cells (40). COUP-TF enhances RAR β expression by increasing the recruitment of receptor coactivators to the RAR β promoter. Stable expression of COUP-TF in MDA-MB-231 cells restored the ability of ATRA to induce RAR β expression (40). Therefore, we examined whether COUP-TF expression was also involved in the induction of RAR β by SR11237 and ciglitazone. Treatment of MDA-MB-231 COUP-TF stable clone cells with ATRA resulted in strong expression of RAR β , as reported previously (40). SR11237 or ciglitazone alone slightly induced RAR β expression in this COUP-TF stable clone (Fig. 8B). As observed previously with the T-47D, ZR-75-1, and Calu-6 cells (Fig. 3), 15d-PGJ $_2$, either alone or in combination with SR11237, did not induce RAR β expression in the MDA-MB-231 COUP-TF stable cells. In contrast, SR11237 and ciglitazone markedly induced RAR β expression (Fig. 8B). Thus, the expression of COUP-TF modulates the induction of RAR β by PPAR γ /RXR heterodimers, as well as the RAR/RXR heterodimers.

DISCUSSION

Retinoids are effective in suppressing tumor development in many animal carcinogenesis models and are being evaluated in clinical trials for prevention and treatment of cancers (23, 24). As an illustration, the rexinoid LGD1069 suppresses mammary tumor development and inhibits the growth of established tumors *in vivo* (57, 58). This retinoid was recently approved for treatment of cutaneous T-cell lymphoma on the basis of clinical trial results and highlights the potential for rexinoids as effective cancer therapeutic agents. Unfortunately, a major limitation in retinoid therapy is that the concentrations needed for anticancer activity also produce adverse effects. Among recent anticancer approaches, combination therapy may lead to synergistic growth-inhibitory effects on cancer cells, thereby allowing the use of lower concentrations to reduce toxicity associated with retinoid treatment (23, 24). In this study, we demonstrate that the PPAR γ ligand ciglitazone and the RXR ligand SR11237 cooperatively inhibited the growth of breast and lung cancer cells, whereas either one alone did not markedly inhibit growth (Fig. 1A). The combination resulted in an enhanced inhibition of cell growth, BrdUrd incorporation, and induction of apoptosis (Fig. 1). Previous studies showed that the rexinoids enhanced the antidiabetic activity of PPAR γ ligands (59). Our present results indicate that the combination of rexinoids and PPAR γ ligands may represent a new approach to effectively inhibit the growth of cancer cells.

The cooperative effect of rexinoids and PPAR γ ligands is likely mediated by their receptors RXR and PPAR γ and their heterodimerization. In elucidation of the downstream pathways mediating the RXR and PPAR γ heterodimers, we observed that induction of RAR β , a potent growth inhibitor (29, 30), is involved in the cooperative growth inhibition of PPAR γ and RXR ligands. Retinoids and ciglitazone alone did not show an appreciable effect on RAR β expression. However, their combination strongly induced RAR β expression in breast cancer and lung cancer cells to a degree that was comparable with that of ATRA (Fig. 2). Thus, our results suggest that induction of RAR β expression accounts for the cooperative growth-inhibitory effect of rexinoids and ciglitazone.

The effect of ciglitazone in inducing RAR β is mediated by PPAR γ . This was demonstrated by our observation that RAR β induction was attenuated by the PPAR γ antagonist BADGE (Fig. 4). Induction of RAR β by ATRA is mainly mediated by RAR/RXR heterodimers that

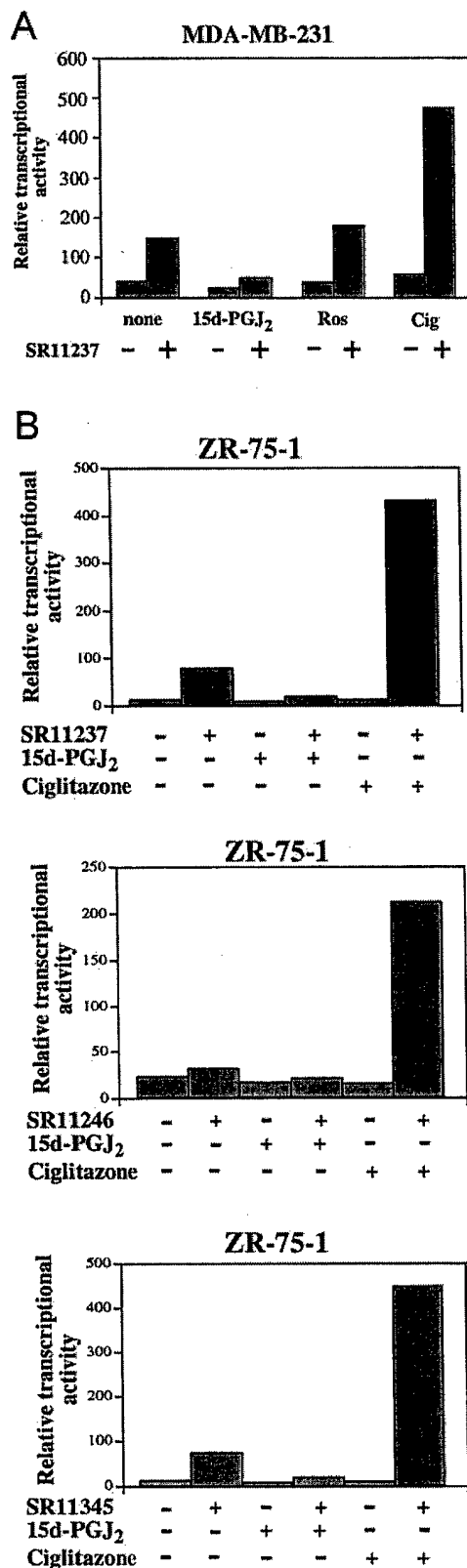


Fig. 6. PPAR γ ligands differentially promote β RARE transcriptional activity. The MDA-MB-231 and ZR-75-1 breast cancer cell lines were transiently transfected with 200 ng of β RARE- κ -CAT (50) and 200 ng of β -gal expression vector. Transfected cells were treated for 20 h with the indicated RXR-selective ligands (1 μ M) or PPAR γ ligands (5 μ M 15d-PGJ₂, 10 μ M ciglitazone, or rosiglitazone), either alone or in combination. Transcriptional activity of β RARE was assessed by CAT assay, using β -gal as an internal standard to evaluate transfection efficiency. Results are representative of three separate experiments, and values are expressed as relative transcriptional activity. A, effect of RXR and PPAR γ ligands on inducing β RARE in MDA-MB-231 cells. B, effect of RXR and PPAR γ ligands on inducing β RARE transcriptional activity in ZR-75-1 cells.

bind to the β RARE in the RAR β promoter (50, 54). In studying how PPAR γ and RXR mediated the RAR β induction by rexinoids and ciglitazone, we demonstrated that the PPAR γ /RXR heterodimeric complex bound to the β RARE (Fig. 5) and induced its transcriptional activation in the presence of rexinoids and ciglitazone (Fig. 6). We demonstrated previously (39, 60) that rexinoids could induce RAR β expression through TR3/RXR heterodimers via β RARE. The results from the present study demonstrate that the PPAR γ /RXR heterodimer represents another RXR-containing heterodimer that mediates the effect of RXR ligands on RAR β induction and growth inhibition.

Classical retinoids fail to induce RAR β expression in certain lung cancer cell lines and in the estrogen-independent MDA-MB-231 breast cancer cells (35, 41–44, 61–64). Lack of RAR β induction has contributed to the retinoid resistance of cancer cells (29, 30). We reported previously (40) that the inability of RAR/RXR heterodimers to activate the RAR β promoter in ATRA-resistant MDA-MB-231 cells was due to lack of the orphan receptor COUP-TF. Our present data indicate that the SR11237 and ciglitazone combination failed to induce RAR β expression in wild-type MDA-MB-231 cells (Fig. 8A)

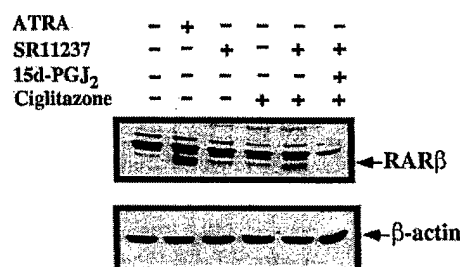


Fig. 7. 15d-PGJ₂ antagonizes RAR β expression induced by rexinoid and ciglitazone. ZR-75-1 cells were treated for 24 h with ATRA alone (1 μ M) or with SR11237 (1 μ M), ciglitazone (10 μ M), or 15d-PGJ₂ (5 μ M), either alone or in combination. RAR β protein expression was determined by Western analysis.

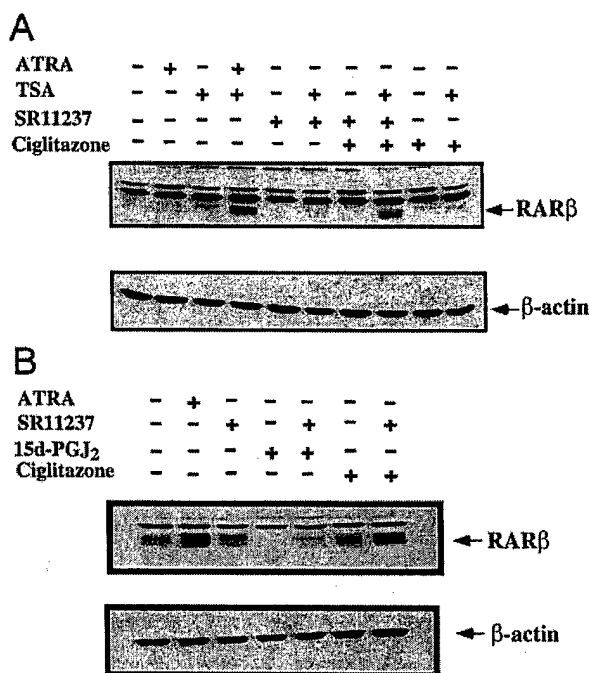


Fig. 8. RXR and PPAR γ ligands differentially regulate RAR β expression in wild-type MDA-MB-231 and MDA-MB-231 COUP-TF stable cells. A, MDA-MB-231 cells were treated for 48 h with SR11237 (1 μ M) ciglitazone (10 μ M), or TSA (100 ng/ml), alone or in combination. RAR β protein expression was determined by Western analysis. B, MDA-MB-231 COUP-TF stable cells were treated for 24 h with SR11237 (1 μ M), 15d-PGJ₂ (5 μ M), or ciglitazone (10 μ M), alone or in combination, and analyzed for RAR β expression by Western analysis.

but strongly induced RAR β in MDA-MB-231 cells stably expressing COUP-TF (Fig. 8B). These results indicate a requirement for COUP-TF in activating RAR β promoter by the PPAR γ /RXR heterodimer. Similar to ATRA, we observed that the combination of ciglitazone and SR11237 strongly induced RAR β in wild-type MDA-MB-231 cells when the HDAC inhibitor TSA was present (Fig. 8A). These results suggest that histone deacetylation is another mechanism responsible for silencing RAR β expression.

One interesting observation in the present study is that PPAR γ ligands differentially regulate β RARE activity and induction of RAR β expression. Ciglitazone, troglitazone, rosiglitazone, and 15d-PGJ₂ act as potent agonists of the PPAR γ /RXR heterodimer on the PPRE (3–8). However, only ciglitazone activated β RARE when used with rexinoids (Fig. 6). Combination of rosiglitazone or 15d-PGJ₂ with a rexinoid failed to induce RAR β in these cancer cell lines (Fig. 3). Thus, different PPAR γ ligands exhibit opposing effects on transactivation of the PPAR γ /RXR heterodimer. Why there is such disparity among the PPAR γ compounds and their ability to cooperate with RXR ligands to induce RAR β is presently unclear. One obvious explanation would be the differences in ligand structure, which may bind in an alternate conformation when bound to PPAR γ /RXR heterodimers complexed with the β RARE. This difference in binding may then impair or fail to initiate efficient transcription, perhaps through inappropriate recruitment of corepressors or coactivators. Regulation of retinoid signaling by receptor polarity and allosteric control of ligand binding has been well demonstrated for RAR/RXR heterodimers. Binding of RAR/RXR heterodimers with RAR ligand strongly activates the DR5 element, whereas the binding suppresses RXR agonist activity on the DR1 element (65, 66). The differential effects of ligands to activate RAR on response elements was shown to result from opposite polarities of the RXR/RAR heterodimer to asymmetrically oriented half-sites (65). Previous studies demonstrated that PPAR γ binds to the 5'-half-site position of the PPRE, whereas RXR occupies the 3'-half site (67, 68). It is likely that similar receptor polarity and allosteric control of transcription of RXR ligand activity by PPAR γ ligand binding exists with respect to the PPRE, which is a DR1 response element, and β RARE, a DR5 element. Whether such an allosteric mechanism exists for the PPAR γ /RXR heterodimer requires further investigation. Regardless of the underlying molecular mechanisms, our observation provides an opportunity to identify specific PPAR γ ligands for inhibiting cancer cell growth through inducing RAR β expression in combination with rexinoids.

In summary, we have demonstrated that rexinoids and ciglitazone can synergistically inhibit the growth of breast and lung cancer cells through their induction of RAR β . Our results demonstrate that PPAR γ /RXR heterodimers can bind to the β RARE and promote its transcriptional activity in response to rexinoids and certain PPAR γ ligands. Further characterization of the effect of PPAR γ ligands on the transactivation of β RARE by PPAR γ /RXR heterodimers and the underlying molecular mechanisms may lead to the identification of potent and specific PPAR γ ligands that inhibit cancer cell growth through this novel signaling pathway.

ACKNOWLEDGMENTS

We thank Laura Frazer for preparation of the manuscript and John Kim for proofreading of the manuscript.

REFERENCES

- Mangelsdorf, D. J., Thummel, C., Beato, M., Herrlich, P., Schutz, G., Umesono, K., Blumberg, B., Kastner, P., Mark, M., Chambon, P., *et al.* The nuclear receptor superfamily: the second decade. *Cell*, 83: 835–839, 1995.
- Moller, D. E., and Berger, J. The mechanisms of action of PPARs. *Annu. Rev. Med.*, 53: 409–435, 2002.
- Rosen, E. D., Walkey, C. J., Puigserver, P., and Spiegelman, B. M. Transcriptional regulation of adipogenesis. *Genes Dev.*, 14: 1293–1307, 2000.
- Rangwala, S. M., and Lazar, M. A. Transcriptional control of adipogenesis. *Annu. Rev. Nutr.*, 20: 535–559, 2000.
- Rosen, E. D., and Spiegelman, B. M. PPAR γ : a nuclear regulator of metabolism, differentiation, and cell growth. *J. Biol. Chem.*, 276: 37731–37734, 2001.
- Hibi, A. K., Michalik, L., and Wahli, W. PPARs: transcriptional effectors of fatty acids and their derivatives. *Cell. Mol. Life Sci.*, 59: 790–798, 2002.
- Sporn, M. B., Suh, N., and Mangelsdorf, D. J. Prospects for prevention and treatment of cancer with selective PPAR γ modulators (SPARMs). *Trends Mol. Med.*, 7: 395–400, 2001.
- Kersten, S., Desvergne, B., and Wahli, W. Roles of PPARs in health and disease. *Nature (Lond.)*, 405: 421–424, 2000.
- Murphy, G. J., and Holder, J. C. PPAR γ agonists: therapeutic role in diabetes, inflammation and cancer. *Trends Pharmacol. Sci.*, 21: 469–474, 2000.
- Clay, C. E., Atsumi, G. I., High, K. P., and Chilton, F. H. Early *de novo* gene expression is required for 15-deoxy- $\Delta^{12,14}$ -prostaglandin I_2 -induced apoptosis in breast cancer cells. *J. Biol. Chem.*, 276: 47131–47135, 2001.
- Kilgore, M. W., Tate, P. L., Rai, S., Sengoku, E., and Price, T. M. MCF-7 and T47D human breast cancer cells contain a functional peroxisomal response. *Mol. Cell. Endocrinol.*, 129: 229–235, 1997.
- Mueller, E., Sarraf, P., Tontonoz, P., Evans, R. M., Martin, K. J., Zhang, M., Fletcher, C., Singer, S., and Spiegelman, B. M. Terminal differentiation of human breast cancer through PPAR γ . *Mol. Cell*, 1: 465–470, 1998.
- Suh, N., Wang, Y., Williams, C. R., Risingsong, R., Gilmer, T., Willson, T. M., and Sporn, M. B. A new ligand for the peroxisome proliferator-activated receptor γ (PPAR γ), GW7845, inhibits rat mammary carcinogenesis. *Cancer Res.*, 59: 5671–5673, 1999.
- Pignatelli, M., Cortes-Canteli, M., Lai, C., Santos, A., and Perez-Castillo, A. The peroxisome proliferator-activated receptor γ is an inhibitor of ErbBs activity in human breast cancer cells. *J. Cell Sci.*, 114: 4117–4126, 2001.
- Thoenes, S. R., Tate, P. L., Price, T. M., and Kilgore, M. W. Differential transcriptional activation of peroxisome proliferator-activated receptor γ by ω -3 and ω -6 fatty acids in MCF-7 cells. *Mol. Cell. Endocrinol.*, 160: 67–73, 2000.
- Yin, F., Wakino, S., Liu, Z., Kim, S., Hsueh, W. A., Collins, A. R., Van Herle, A. J., and Law, R. E. Troglitazone inhibits growth of MCF-7 breast carcinoma cells by targeting G₁ cell cycle regulators. *Biochem. Biophys. Res. Commun.*, 286: 916–922, 2001.
- Elstner, E., Muller, C., Koshizuka, K., Williamson, E. A., Park, D., Asou, H., Shintaku, P., Said, J. W., Heber, D., and Koeffler, H. P. Ligands for peroxisome proliferator-activated receptor γ and retinoic acid receptor inhibit growth and induce apoptosis of human breast cancer cells *in vitro* and in BNX mice. *Proc. Natl. Acad. Sci. USA*, 95: 8806–8811, 1998.
- Mehta, R. G., Williamson, E., Patel, M. K., and Koeffler, H. P. A ligand of peroxisome proliferator-activated receptor γ , retinoids, and prevention of preneoplastic mammary lesions. *J. Natl. Cancer Inst. (Bethesda)*, 92: 418–423, 2000.
- Theocharis, S., Kanelli, H., Politi, E., Margeli, A., Karkandaris, C., Philippides, T., and Koutselis, A. Expression of peroxisome proliferator activated receptor γ in non-small cell lung carcinoma: correlation with histological type and grade. *Lung Cancer*, 36: 249–255, 2002.
- Satoh, T., Toyoda, M., Hoshino, H., Monden, T., Yamada, M., Shimizu, H., Miyamoto, K., and Mori, M. Activation of peroxisome proliferator-activated receptor γ stimulates the growth arrest and DNA-damage inducible 153 gene in non-small cell lung carcinoma cells. *Oncogene*, 21: 2171–2180, 2002.
- Tsubouchi, Y., Sano, H., Kawahito, Y., Mukai, S., Yamada, R., Kohno, M., Inoue, K., Hla, T., and Kondo, M. Inhibition of human lung cancer cell growth by the peroxisome proliferator-activated receptor γ agonists through induction of apoptosis. *Biochem. Biophys. Res. Commun.*, 270: 400–405, 2000.
- Chang, T. H., and Szabo, E. Induction of differentiation and apoptosis by ligands of peroxisome proliferator-activated receptor γ in non-small cell lung cancer. *Cancer Res.*, 60: 1129–1138, 2000.
- Lippman, S. M., and Lotan, R. Advances in the development of retinoids as chemopreventive agents. *J. Nutr.*, 130: 479S–482S, 2000.
- Altucci, L., and Gronemeyer, H. The promise of retinoids to fight against cancer. *Nat. Rev. Cancer*, 1: 181–193, 2001.
- Chambon, P. A decade of molecular biology of retinoic acid receptors. *FASEB J.*, 10: 940–954, 1996.
- Zhang, X. K., Hoffmann, B., Tran, P. B., Graupner, G., and Pfahl, M. Retinoid X receptor is an auxiliary protein for thyroid hormone and retinoic acid receptors. *Nature (Lond.)*, 355: 441–446, 1992.
- Nunez, S. B., Medin, J. A., Braissant, O., Kemp, L., Wahli, W., Ozato, K., and Segars, J. H. Retinoid X receptor and peroxisome proliferator-activated receptor activate an estrogen responsive gene independent of the estrogen receptor. *Mol. Cell. Endocrinol.*, 127: 27–40, 1997.
- Stoll, B. A. Linkage between retinoid and fatty acid receptors: implications for breast cancer prevention. *Eur. J. Cancer Prev.*, 11: 319–325, 2002.
- Zhang, X. K. Vitamin A and apoptosis in prostate cancer. *Endocr. Relat. Cancer*, 9: 87–102, 2002.
- Zhang, X. K., Liu, Y., and Lee, M. O. Retinoid receptors in human lung cancer and breast cancer. *Mutat. Res.*, 350: 267–277, 1996.
- Gebert, J. F., Moghal, N., Frangioni, J. V., Sugarbaker, D. J., and Neel, B. G. High frequency of retinoic acid receptor β abnormalities in human lung cancer. *Oncogene*, 6: 1859–1868, 1991.

32. Houle, B., Leduc, F., and Bradley, W. E. Implication of RAR β in epidermoid (squamous) lung cancer. *Genes Chromosomes Cancer*, 3: 358–366, 1991.
33. Hu, L., Crowe, D. L., Rheinwald, J. G., Chambon, P., and Gudas, L. J. Abnormal expression of retinoic acid receptors and keratin 19 by human oral and epidermal squamous cell carcinoma cell lines. *Cancer Res.*, 51: 3972–3981, 1991.
34. Nervi, C., Vollberg, T. M., George, M. D., Zelent, A., Chambon, P., and Jetten, A. M. Expression of nuclear retinoic acid receptors in normal tracheobronchial cells and in lung carcinoma cells. *Exp. Cell Res.*, 195: 163–170, 1991.
35. Liu, Y., Lee, M. O., Wang, H. G., Li, Y., Hashimoto, Y., Klaus, M., Reed, J. C., and Zhang, X. Retinoic acid receptor β mediates the growth-inhibitory effect of retinoic acid by promoting apoptosis in human breast cancer cells. *Mol. Cell. Biol.*, 16: 1138–1149, 1996.
36. Swisshelm, K., Ryan, K., Lee, X., Tsou, H. C., Peacocke, M., and Sager, R. Down-regulation of retinoic acid receptor β in mammary carcinoma cell lines and its up-regulation in senescing normal mammary epithelial cells. *Cell Growth Differ.*, 5: 133–141, 1994.
37. Berard, J., Laboune, F., Mukuna, M., Masse, S., Kothary, R., and Bradley, W. E. Lung tumors in mice expressing an antisense RAR β 2 transgene. *FASEB J.*, 10: 1091–1097, 1996.
38. Lotan, R., Xu, X. C., Lippman, S. M., Ro, J. Y., Lee, J. S., Lee, J. J., and Hong, W. K. Suppression of retinoic acid receptor β in premalignant oral lesions and its up-regulation by isotretinoin. *N. Engl. J. Med.*, 332: 1405–1410, 1995.
39. Wu, Q., Dawson, M. I., Zheng, Y., Hobbs, P. D., Agadir, A., Jong, L., Li, Y., Liu, R., Lin, B., and Zhang, X. K. Inhibition of *trans*-retinoic acid-resistant human breast cancer cell growth by retinoid X receptor-selective retinoids. *Mol. Cell. Biol.*, 17: 6598–6608, 1997.
40. Lin, B., Chen, G. Q., Xiao, D., Kolluri, S. K., Cao, X., Su, H., and Zhang, X. K. Orphan receptor COUP-TF is required for induction of retinoic acid receptor β , growth inhibition, and apoptosis by retinoic acid in cancer cells. *Mol. Cell. Biol.*, 20: 957–970, 2000.
41. Bovenzi, V., and Momparler, R. L. Antineoplastic action of 5-aza-2'-deoxycytidine and histone deacetylase inhibitor and their effect on the expression of retinoic acid receptor β and estrogen receptor α genes in breast carcinoma cells. *Cancer Chemother. Pharmacol.*, 48: 71–76, 2001.
42. Sirchia, S. M., Ferguson, A. T., Sironi, E., Subramanyan, S., Orlandi, R., Sukumar, S., and Sacchi, N. Evidence of epigenetic changes affecting the chromatin state of the retinoic acid receptor β 2 promoter in breast cancer cells. *Oncogene*, 19: 1556–1563, 2000.
43. Widschwendter, M., Berger, J., Hermann, M., Muller, H. M., Amberger, A., Zeschmick, M., Widschwendter, A., Abendstein, B., Zeimet, A. G., Daxenbichler, G., and Marth, C. Methylation and silencing of the retinoic acid receptor β 2 gene in breast cancer. *J. Natl. Cancer Inst. (Bethesda)*, 92: 826–832, 2000.
44. Sirchia, S. M., Ren, M., Pili, R., Sironi, E., Somenzi, G., Ghidoni, R., Toma, S., Nicolo, G., and Sacchi, N. Endogenous reactivation of the RAR β 2 tumor suppressor gene epigenetically silenced in breast cancer. *Cancer Res.*, 62: 2455–2461, 2002.
45. Momparler, R. L., and Bovenzi, V. DNA methylation and cancer. *J. Cell Physiol.*, 183: 145–154, 2000.
46. Wright, H. M., Clish, C. B., Mikami, T., Hauser, S., Yanagi, K., Hiramatsu, R., Serhan, C. N., and Spiegelman, B. M. A synthetic antagonist for the peroxisome proliferator-activated receptor γ inhibits adipocyte differentiation. *J. Biol. Chem.*, 275: 1873–1877, 2000.
47. Lehmann, J. M., Jong, L., Fanjul, A., Cameron, J. F., Lu, X. P., Haefner, P., Dawson, M. I., and Pfahl, M. Retinoids selective for retinoid X receptor response pathways. *Science (Wash. DC)*, 258: 1944–1946, 1992.
48. Dawson, M. I., Jong, L., Hobbs, P. D., Cameron, J. F., Chao, W. R., Pfahl, M., Lee, M. O., Shroot, B., and Pfahl, M. Conformational effects of retinoid receptor selectivity. 2. Effects of retinoid bridging group on retinoid X receptor activity and selectivity. *J. Med. Chem.*, 38: 3368–3383, 1995.
49. Dawson, M. I., Hobbs, P. D., Jong, L., Xiao, D., Chao, W. R., Pan, C., and Zhang, X. K. sp²-bridged diaryl retinoids: effects of bridge-region substitution on retinoid X receptor selectivity. *Bioorg. Med. Chem. Lett.*, 10: 1307–1310, 2000.
50. Hoffmann, B., Lehmann, J. M., Zhang, X. K., Hermann, T., Husmann, M., Graupner, G., and Pfahl, M. A retinoic acid receptor-specific element controls the retinoic acid receptor- β promoter. *Mol. Endocrinol.*, 4: 1727–1736, 1990.
51. James, S. Y., Mercer, E., Brady, M., Binderup, L., and Colston, K. W. EB1089, a synthetic analogue of vitamin D, induces apoptosis in breast cancer cells *in vivo* and *in vitro*. *Br. J. Pharmacol.*, 125: 953–962, 1998.
52. Mangelsdorf, D. J., and Evans, R. M. The RXR heterodimers and orphan receptors. *Cell*, 83: 841–850, 1995.
53. Li, Y., Dawson, M. I., Agadir, A., Lee, M. O., Jong, L., Hobbs, P. D., and Zhang, X. K. Regulation of RAR β expression by RAR- and RXR-selective retinoids in human lung cancer cell lines: effect on growth inhibition and apoptosis induction. *Int. J. Cancer*, 75: 88–95, 1998.
54. Sucov, H. M., Murakami, K. K., and Evans, R. M. Characterization of an autoregulated response element in the mouse retinoic acid receptor type β gene. *Proc. Natl. Acad. Sci. USA*, 87: 5392–5396, 1990.
55. Fontana, J. A., Hobbs, P. D., and Dawson, M. I. Inhibition of mammary carcinoma growth by retinoid benzoic acid derivatives. *Exp. Cell Biol.*, 56: 254–263, 1988.
56. van der Burg, B., van der Leede, B. M., Kwakkenbos-Isbrucker, L., Salverda, S., de Laat, S. W., and van der Saag, P. T. Retinoic acid resistance of estradiol-independent breast cancer cells coincides with diminished retinoic acid receptor function. *Mol. Cell. Endocrinol.*, 91: 149–157, 1993.
57. Gottardis, M. M., Bischoff, E. D., Shirley, M. A., Wagoner, M. A., Lamph, W. W., and Heyman, R. A. Chemoprevention of mammary carcinoma by LGD1069 (Targretin): an RXR-selective ligand. *Cancer Res.*, 56: 5566–5570, 1996.
58. Bischoff, E. D., Gottardis, M. M., Moon, T. E., Heyman, R. A., and Lamph, W. W. Beyond tamoxifen: the retinoid X receptor-selective ligand LGD1069 (TARGRETIN) causes complete regression of mammary carcinoma. *Cancer Res.*, 58: 479–484, 1998.
59. Mukherjee, R., Davies, P. J., Crombie, D. L., Bischoff, E. D., Cesario, R. M., Jow, L., Hamann, L. G., Boehm, M. F., Mondon, C. E., Nadzan, A. M., Paterniti, J. R., Jr., and Heyman, R. A. Sensitization of diabetic and obese mice to insulin by retinoid X receptor agonists. *Nature (Lond.)*, 386: 407–410, 1997.
60. Chen, G. Q., Lin, B., Dawson, M. I., and Zhang, X. K. Nicotine modulates the effects of retinoids on growth inhibition and RAR β expression in lung cancer cells. *Int. J. Cancer*, 99: 171–178, 2002.
61. Kim, Y. H., Dohi, D. F., Han, G. R., Zou, C. P., Oridate, N., Walsh, G. L., Nesbitt, J. C., Xu, X. C., Hong, W. K., Lotan, R., *et al.* Retinoid refractoriness occurs during lung carcinogenesis despite functional retinoid receptors. *Cancer Res.*, 55: 5603–5610, 1995.
62. Zhang, X. K., Liu, Y., Lee, M. O., and Pfahl, M. A specific defect in the retinoic acid response associated with human lung cancer cell lines. *Cancer Res.*, 54: 5663–5669, 1994.
63. van der Leede, B. M., van den Brink, C. E., and van der Saag, P. T. Retinoic acid receptor and retinoid X receptor expression in retinoic acid-resistant human tumor cell lines. *Mol. Carcinog.*, 8: 112–122, 1993.
64. Moghal, N., and Neel, B. G. Evidence for impaired retinoic acid receptor-thyroid hormone receptor AF-2 cofactor activity in human lung cancer. *Mol. Cell. Biol.*, 15: 3945–3959, 1995.
65. Kurokawa, R., DiRenzo, J., Boehm, M., Sugarman, J., Gloss, B., Rosenfeld, M. G., Heyman, R. A., and Glass, C. K. Regulation of retinoid signalling by receptor polarity and allosteric control of ligand binding. *Nature (Lond.)*, 371: 528–531, 1994.
66. Zechel, C., Shen, X. Q., Chen, J. Y., Chen, Z. P., Chambon, P., and Gronemeyer, H. The dimerization interfaces formed between the DNA binding domains of RXR, RAR and TR determine the binding specificity and polarity of the full-length receptors to direct repeats. *EMBO J.*, 13: 1425–1433, 1994.
67. Upenberg, A., Jeannin, E., Wahli, W., and Desvergne, B. Polarity and specific sequence requirements of peroxisome proliferator-activated receptor (PPAR)/retinoid X receptor heterodimer binding to DNA. A functional analysis of the malic enzyme gene PPAR response element. *J. Biol. Chem.*, 272: 20108–20117, 1997.
68. Hsu, M. H., Palmer, C. N., Song, W., Griffin, K. J., and Johnson, E. F. A carboxyl-terminal extension of the zinc finger domain contributes to the specificity and polarity of peroxisome proliferator-activated receptor DNA binding. *J. Biol. Chem.*, 273: 27988–27997, 1998.